

Abwasser und resistente Keime: Kann das überwacht werden?

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Relevanz

RICHTLINIE (EU) 2024/3019 DES EUROPÄISCHEN PARLAMENTS UND DES RATES

vom 27. November 2024

über die Behandlung von kommunalem Abwasser

(Neufassung)

Artikel 17

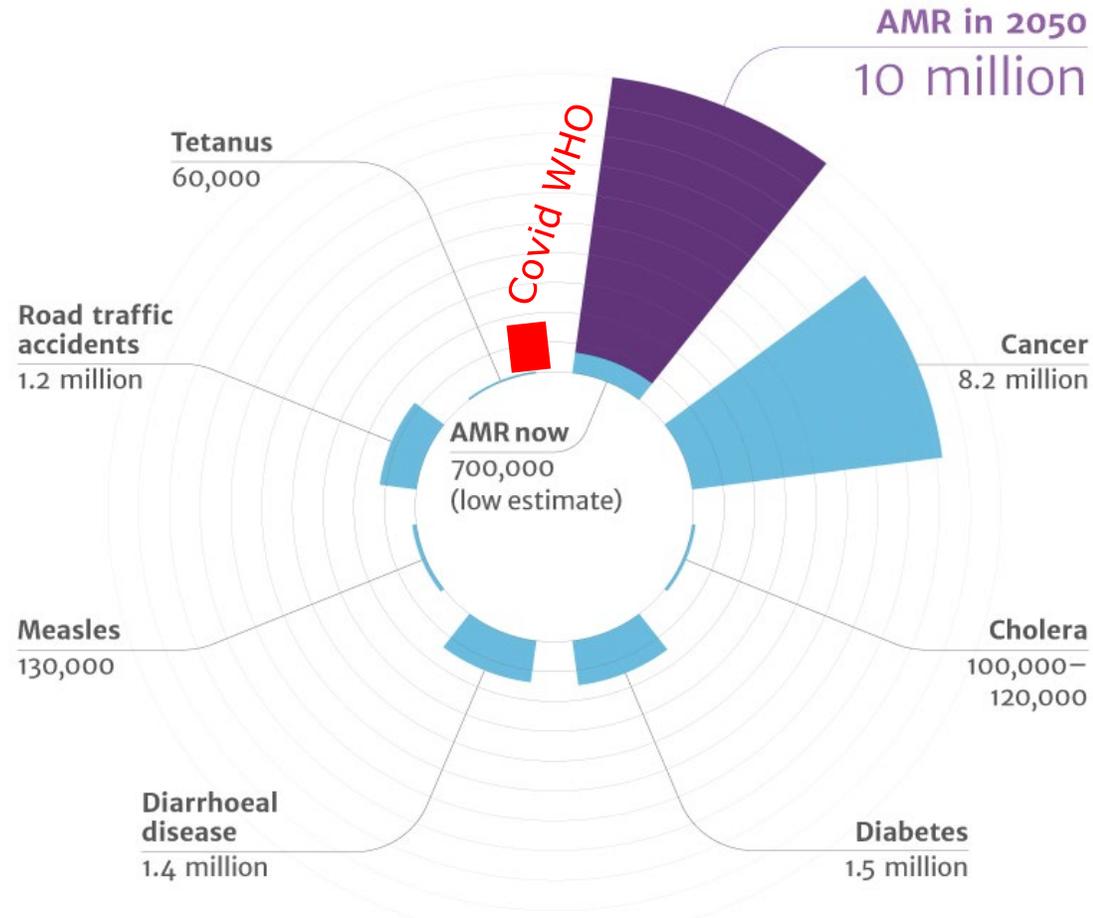
Überwachung von kommunalem Abwasser

(3) Für Siedlungsgebiete mit 100 000 EW und mehr stellen die Mitgliedstaaten bis zum letzten Tag des zweiten Jahres nach Erlass des Durchführungsrechtsakts im Sinne von Unterabsatz 2 sicher, dass antimikrobielle Resistenzen im kommunalen Abwasser überwacht werden.

Zur Erweiterung des Wissens über die wichtigsten Quellen antimikrobieller Resistenzen sollte eine Verpflichtung zur Überwachung des Vorhandenseins antimikrobieller Resistenzen im kommunalen Abwasser eingeführt werden, um unsere wissenschaftlichen Erkenntnisse auszubauen und möglicherweise in Zukunft angemessene Maßnahmen zu ergreifen.

Prognose Anzahl Sterbefälle durch antibiotikaresistente Erreger (AMR) pro Jahr

Problembeschreibung



Sources:

Diabetes: www.who.int/mediacentre/factsheets/fs312/en/ Cancer: www.who.int/mediacentre/factsheets/fs297/en/
Cholera: www.who.int/mediacentre/factsheets/fs107/en/ Diarrhoeal disease: www.sciencedirect.com/science/article/pii/S0140673612617280
Measles: www.sciencedirect.com/science/article/pii/S0140673612617280 Road traffic accidents: www.who.int/mediacentre/factsheets/fs358/en/
Tetanus: www.sciencedirect.com/science/article/pii/S0140673612617280

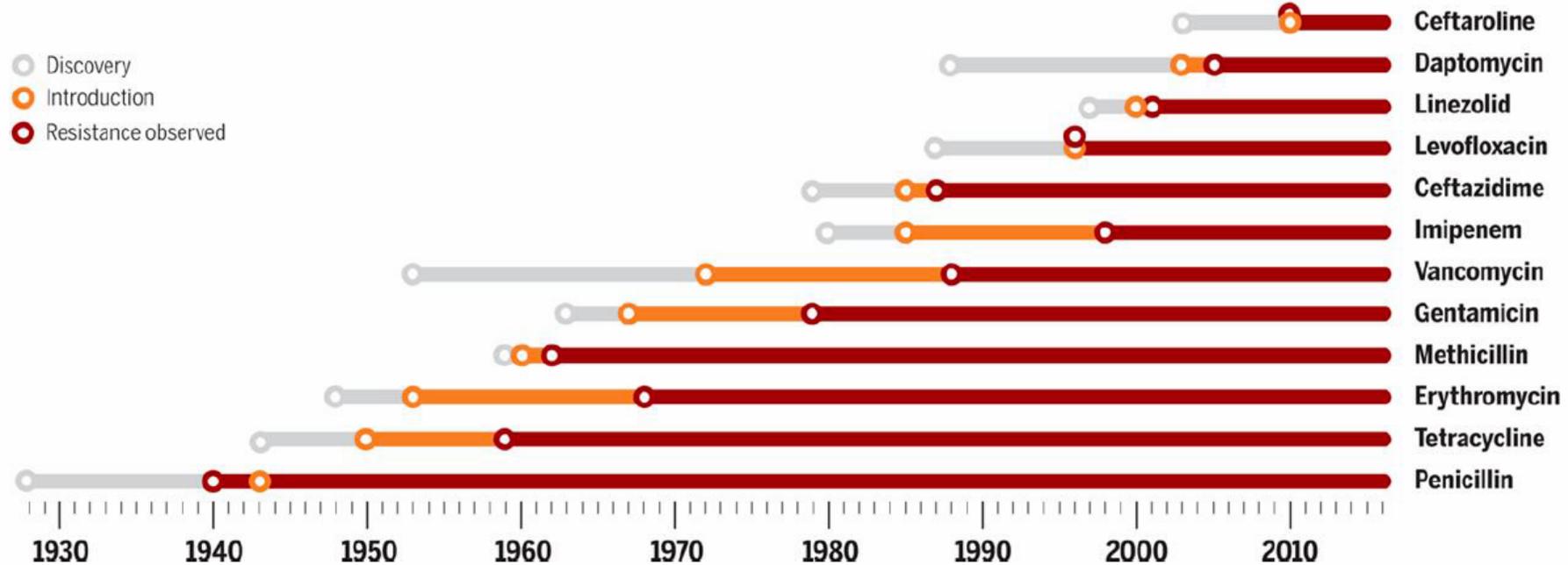


Zeitspanne zwischen Einführung der Antibiotika und der Beobachtung erster Resistenzen

„Die Stille Pandemie“

The rise of resistance

Bacteria have developed resistance to every antibiotic discovered so far, sometimes even before the drug reached the market. The appearance of resistance does not mean that a drug has become completely useless.



Kai Kupferschmidt Science 2016;352:758-761



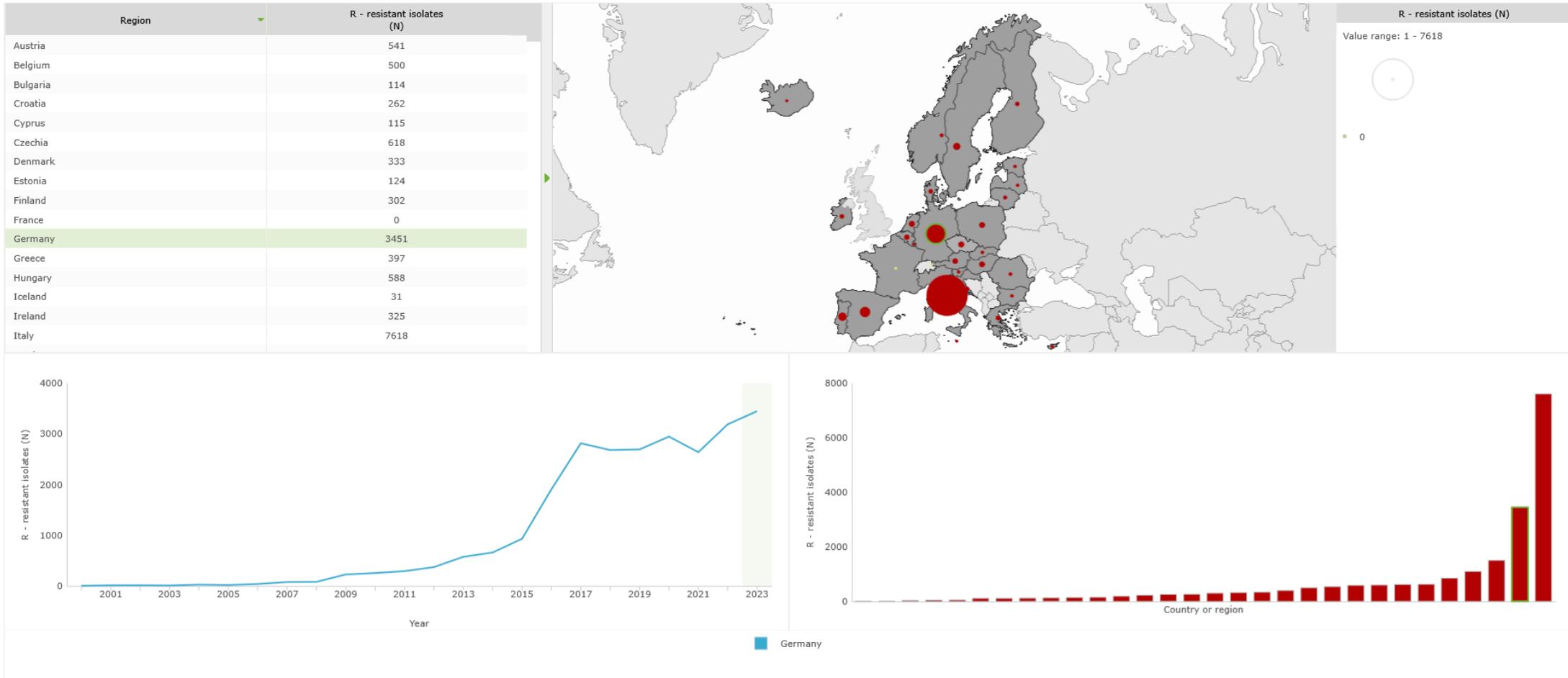
Problembeschreibung

Europäische Datenbank zu klinischen Resistenzen



Surveillance Atlas of Infectious Diseases

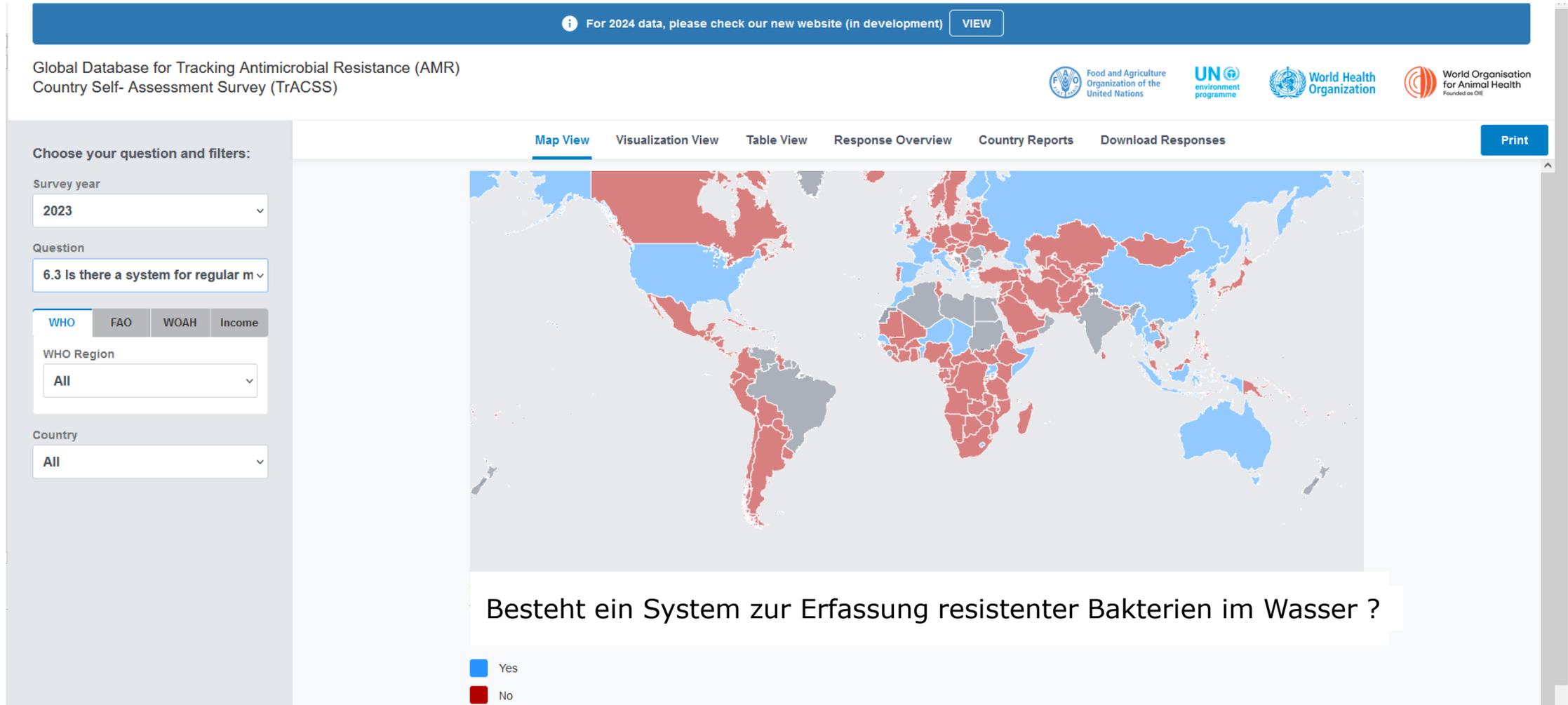
Antimicrobial resistance | Escherichia coli | Third-generation cephalosporins | R - resistant isolates | 2023



Problembeschreibung

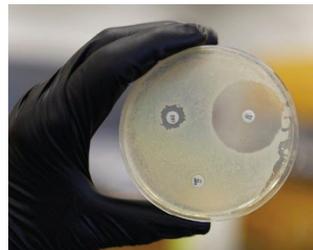
Weltweite Datenbank zur Nachverfolgung von Antibiotikaresistenzen

Problembeschreibung

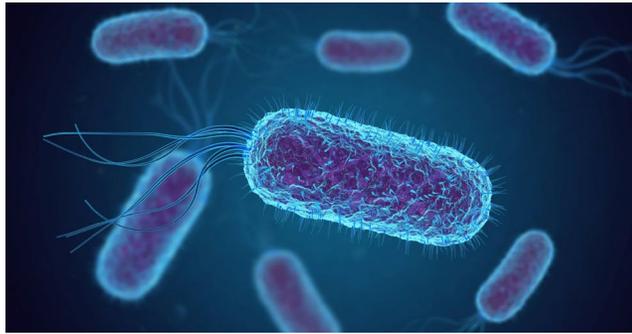


Abwasser Epidemiologie - Monitoring der Resistenzen im Abwasser

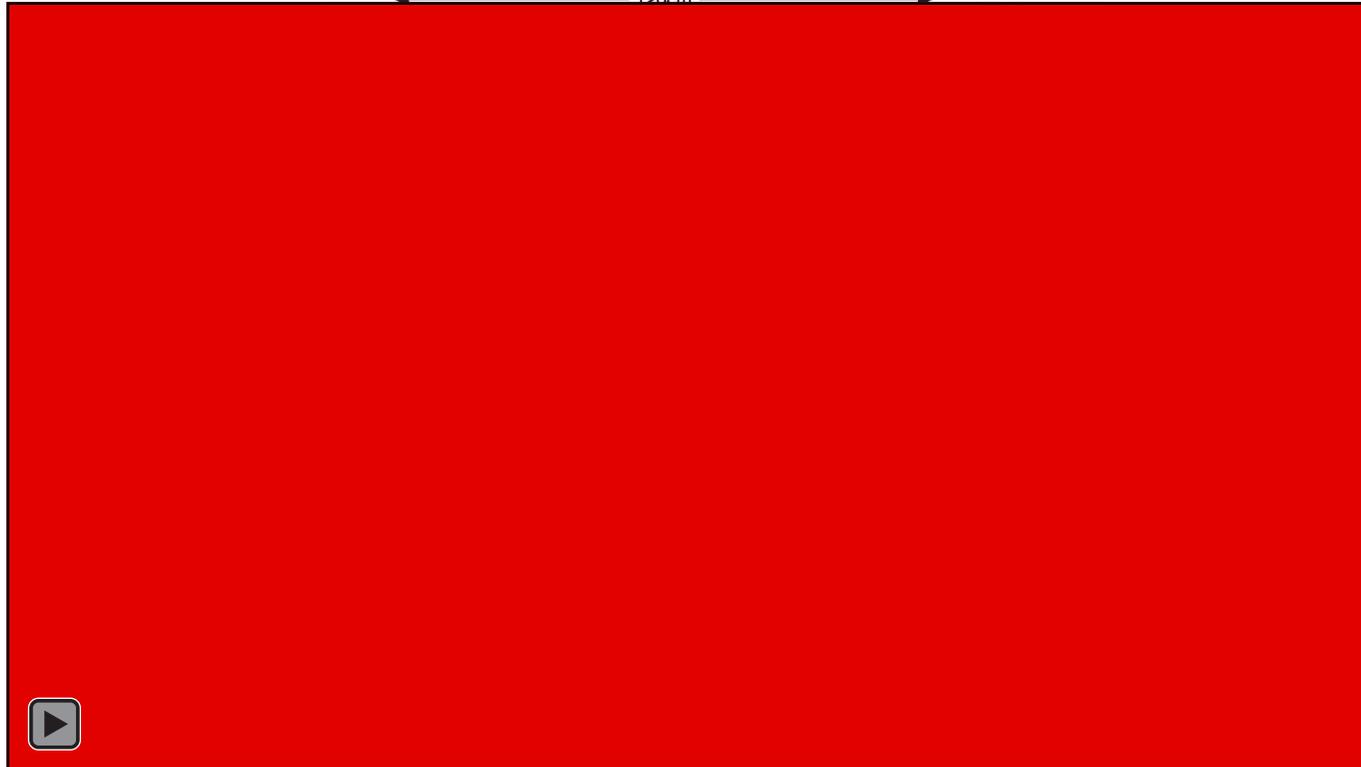
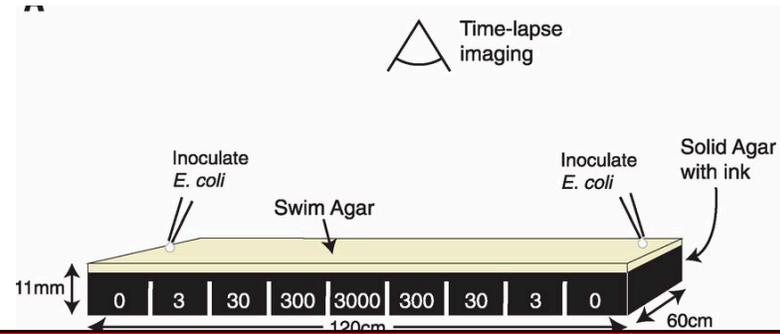
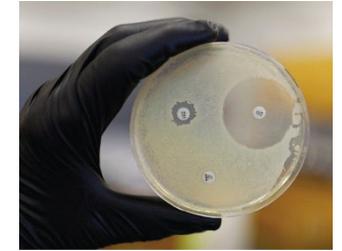
- Virus monitoring ist als Konzept vergleichsweise "einfach"
 - ⇒ Covid Viren zeigen im Abwasser bzw. der Umwelt keine Dynamik
- Antibiotika Resistenz zeigt eine wesentlich komplexere Dynamik
- Epidemiologische Studien beruhen im Wesentlichen auf kulturbasierten Ansätzen
- Aber nur 10% der Umweltbakterien können kultiviert werden



Berendonk et al. Nature Microbiology Review 2015

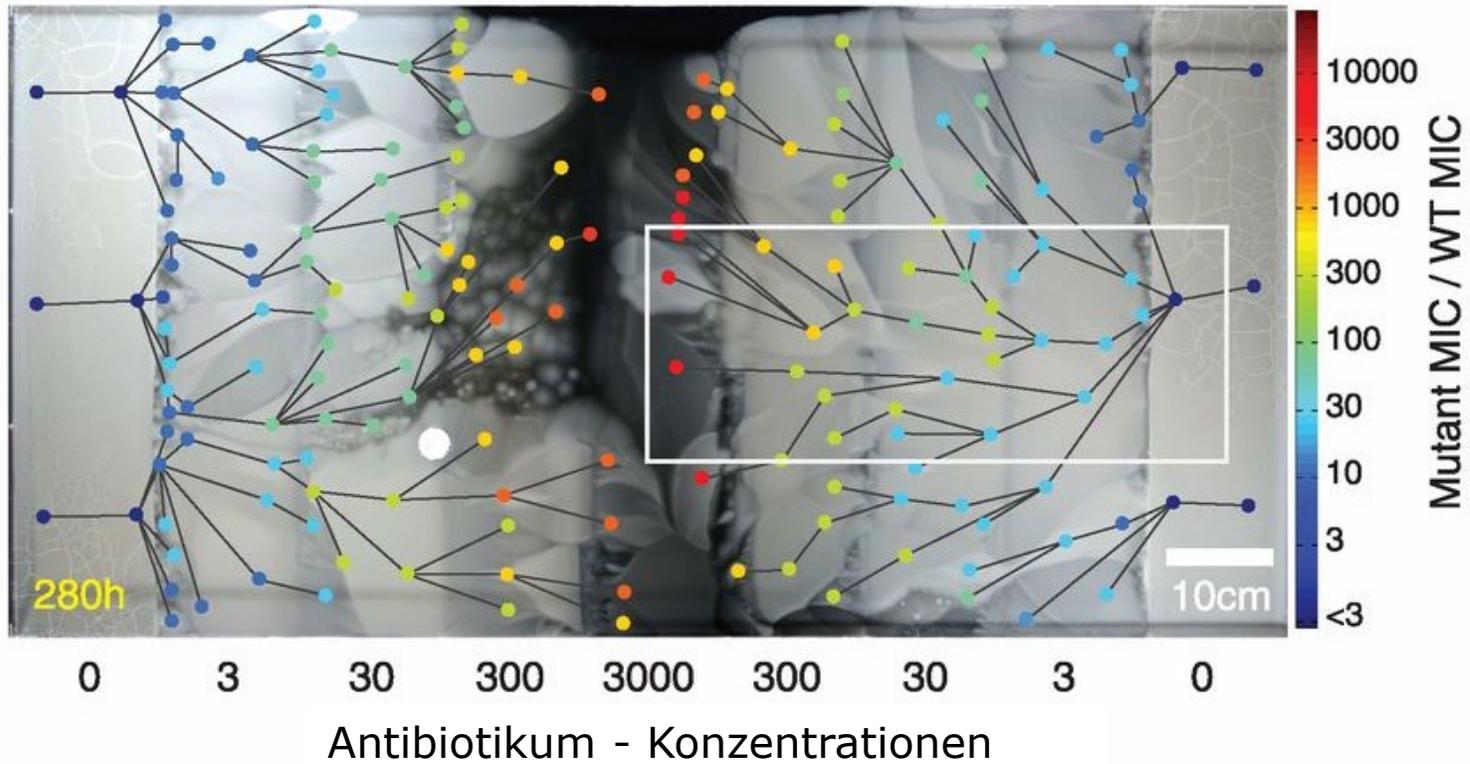


Evolution eines Bakteriums (*E. coli*)



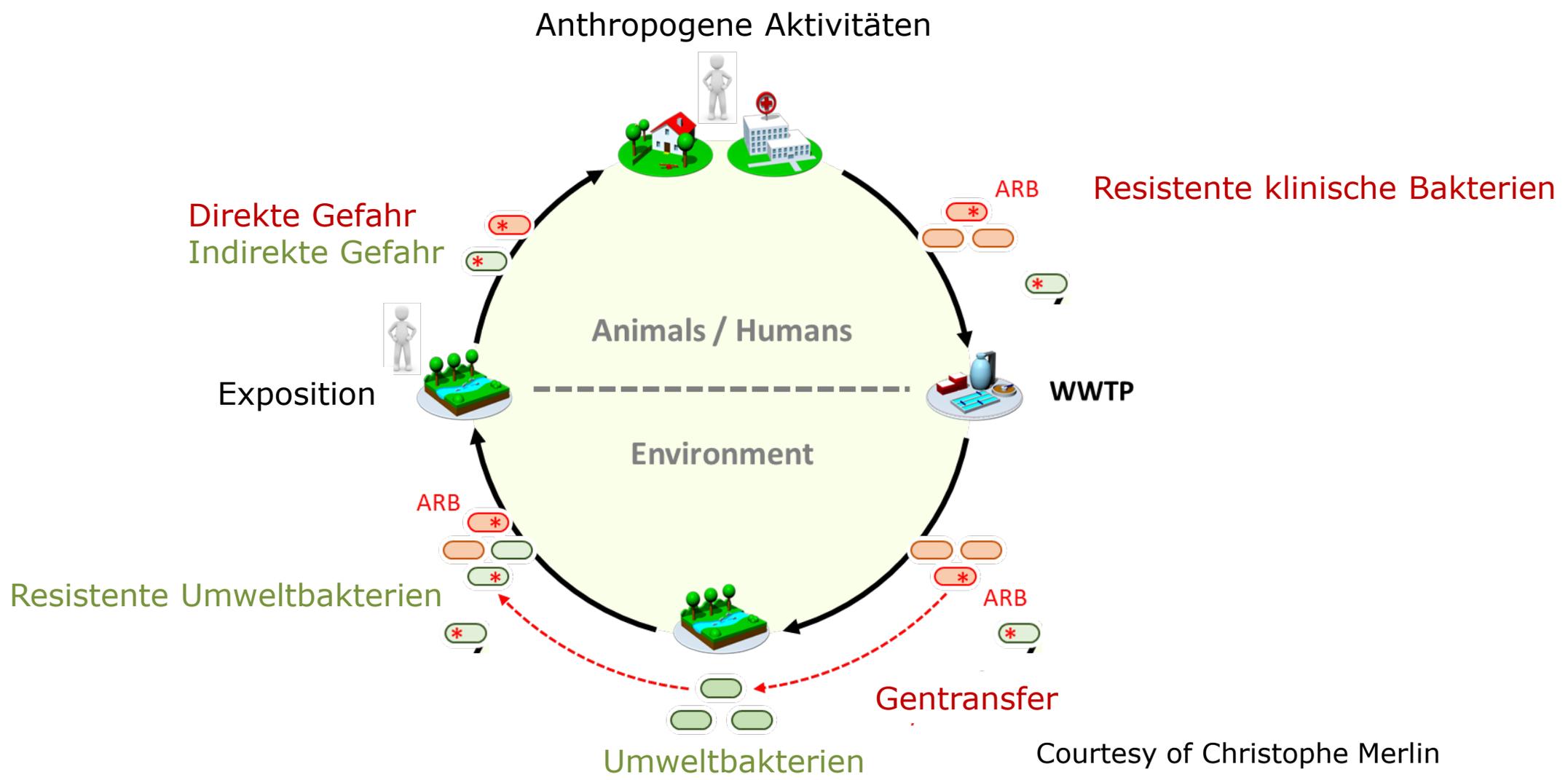
Michael Baym et al. Science 2016;353:1147-1151

Evolution eines Bakteriums (*E. coli*)



Michael Baym et al. Science 2016;353:1147-1151

Resistente Bakterien und Gentransfer



Courtesy of Christophe Merlin

Mögliche Methoden des AMR monitoring

Problembeschreibung

JOURNAL ARTICLE

Antibiotic resistant bacteria and resistance genes in the bottom sediment of a small stream and the potential impact of remobilization ^{FREE}

S Heß , T U Berendonk, D Kneis

FEMS Microbiology Ecology, Volume 94, Issue 9, September 2018, fiy128,
<https://doi.org/10.1093/femsec/fiy128>

Published: 30 June 2018 [Article history](#) ▼

SCIENTIFIC REPORTS

OPEN

High genomic diversity of multi-drug resistant wastewater *Escherichia coli*

Norhan Mahfouz¹, Serena Caucci^{2,3}, Eric Achatz¹, Torsten Semmler^{4*}, Sebastian Guenther^{4,5}, Thomas U. Berendonk² & Michael Schroeder¹

Received: 27 November 2017
 Accepted: 18 May 2018

Water Research
 Volume 162, 1 October 2019, Pages 320-330

Antibiotic resistance genes in treated wastewater and in the receiving water bodies: A pan-European survey of urban settings

Damiano Cacace^a, Despo Fatta-Kassinos^b, Celia M. Manaia^c, Eddie Cytryn^d, Norbert Kreuzinger^e, Luigi Rizzo^f, Papi Karaolia^b, Thomas Schwartz^g, Johannes Alexander^g, Christophe Merlin^h, Hmda Garelickⁱ, Heike Schmitt^j, Daisy de Vries^k, Carsten U. Schwermer^l, Sureyya Meric^l, Can Burak Ozkal^l, Marie-Noelle Pons^m, David Kneis^a, Thomas U. Berendonk^a  

Sewage from Airplanes Exhibits High Abundance and Diversity of Antibiotic Resistance Genes

Stefanie Heß*, David Kneis, Tobias Österlund, Bing Li, Erik Kristiansson, and Thomas U. Berendonk

● **Cite this:** *Environ. Sci. Technol.* 2019, 53, 23, 13898–13905
 Publication Date: November 12, 2019 ▼
<https://doi.org/10.1021/acs.est.9b03236>

Article Views	Altmetric	Citations
2348	106	31

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FEMS MICROBIOLOGY ECOLOGY
 Leading research in the microbial ecology community

FEMS Microbiol Ecol. 2023 Apr; 99(4): fiad031. PMID: PMC10054912
 Published online 2023 Mar 20. doi: [10.1093/femsec/fiad031](https://doi.org/10.1093/femsec/fiad031) PMID: [36941120](https://pubmed.ncbi.nlm.nih.gov/36941120/)

Quantification of the mobility potential of antibiotic resistance genes through multiplexed ddPCR linkage analysis

Magali de la Cruz Barron^{1*}, David Kneis¹, Alan Xavier Elena¹, Kenyum Bagra¹, Thomas U. Berendonk¹, and Uli Klümper^{2*}

Antibiotic Resistance Genes in River Biofilms: A Metagenomic Approach toward the Identification of Sources and Candidate Hosts

David Kneis*, Thomas U. Berendonk, Sofia K. Forslund, and Stefanie Hess

● **Cite this:** *Environ. Sci. Technol.* 2022, 56, 21, 14913–14922
 Publication Date: April 25, 2022 ▼
<https://doi.org/10.1021/acs.est.2c00370>

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Science of The Total Environment
 Volume 904, 15 December 2023, 166661

Environmental stress increases the invasion success of antimicrobial resistant bacteria in river microbial communities

Kenyum Bagra^{a,b}, Xavier Bellanger^c, Christophe Merlin^c, Gargi Singh^b, Thomas U. Berendonk^a, Uli Klümper^a  

Environment International 144 (2020) 106035

Contents lists available at ScienceDirect

Environment International

journal homepage: www.elsevier.com/locate/envint

A global multinational survey of cefotaxime-resistant coliforms in urban wastewater treatment plants

Roberto B.M. Marano^{a,b,1}, Telma Fernandes^{c,1}, Célia M. Manaia^c, Olga Nunes^d, Donald Morrison^x, Thomas U. Berendonk^e, Norbert Kreuzinger^f, Tanel Tenson^g, Gianluca Corno^h,



BRIEF RESEARCH REPORT article
 Front. Public Health. 27 December 2023
 Sec. Infectious Diseases: Epidemiology and Prevention
 Volume 11 - 2023 | <https://doi.org/10.3389/fpubh.2023.1271594>

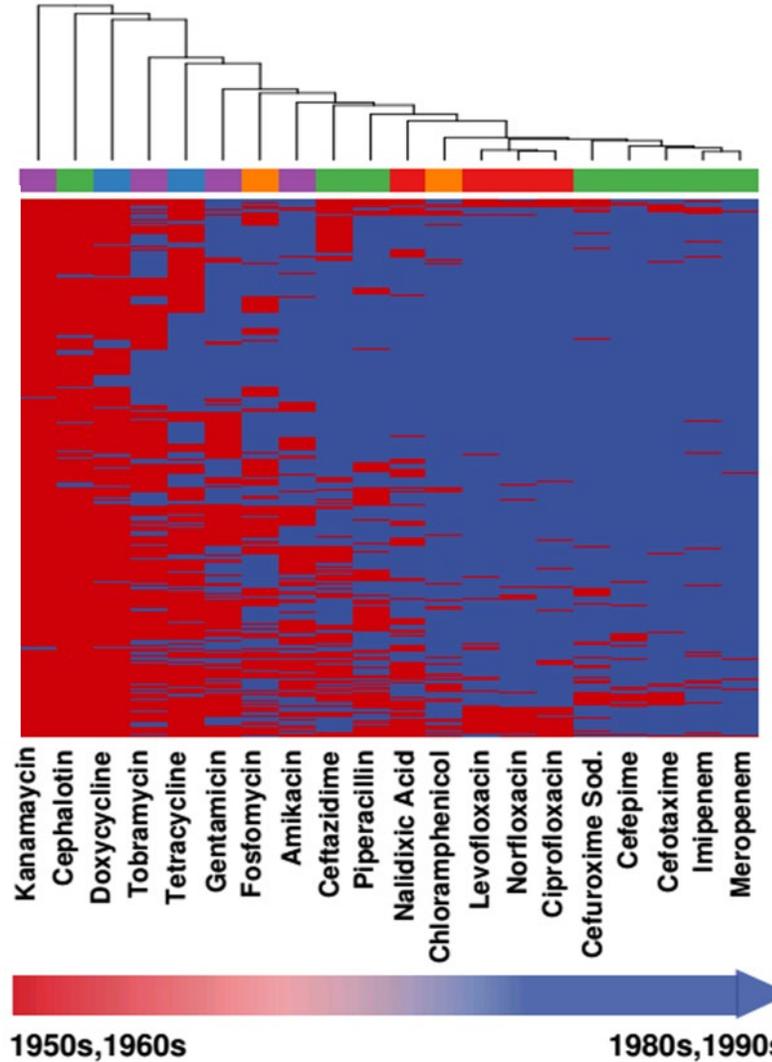
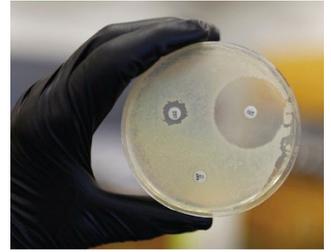
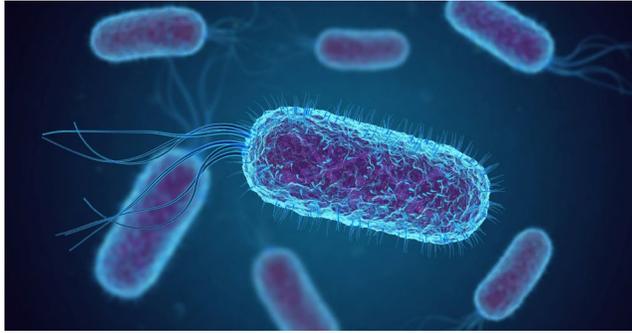
Evaluating the sensitivity of droplet digital PCR for the quantification of SARS-CoV-2 in wastewater

 Magali de la Cruz Barron^{1*}  David Kneis¹  Michael Geissler²  Roger Dumke³
 Alexander Dalpke^{2,3}  Thomas U. Berendonk^{1*}

Beispiel



1100 Antibiotikaresistente individuelle Bakterien (E. coli) des Abwassers

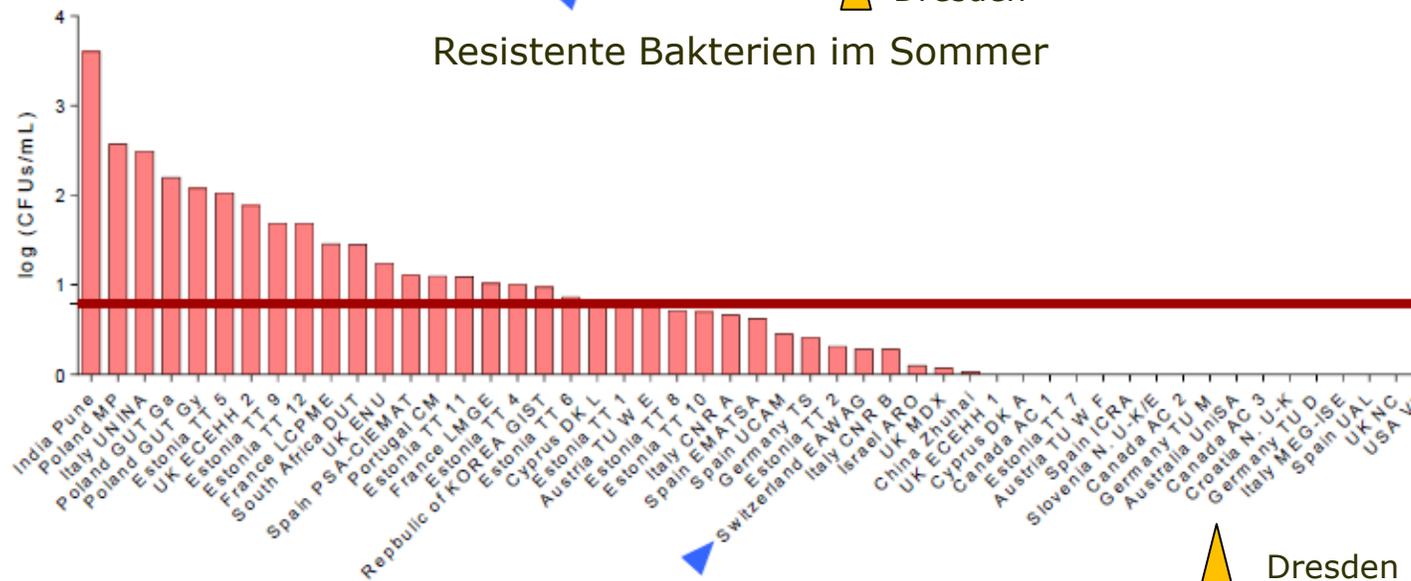
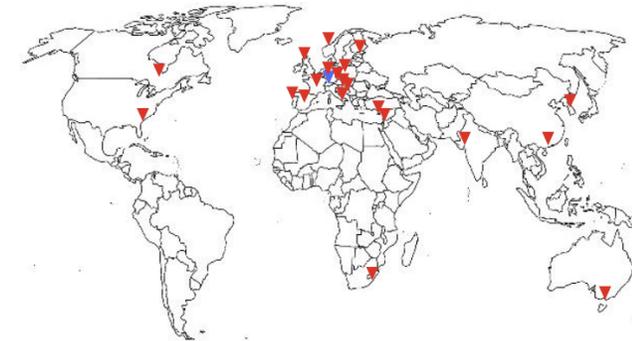
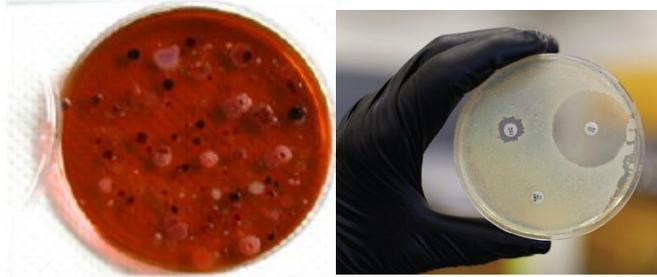
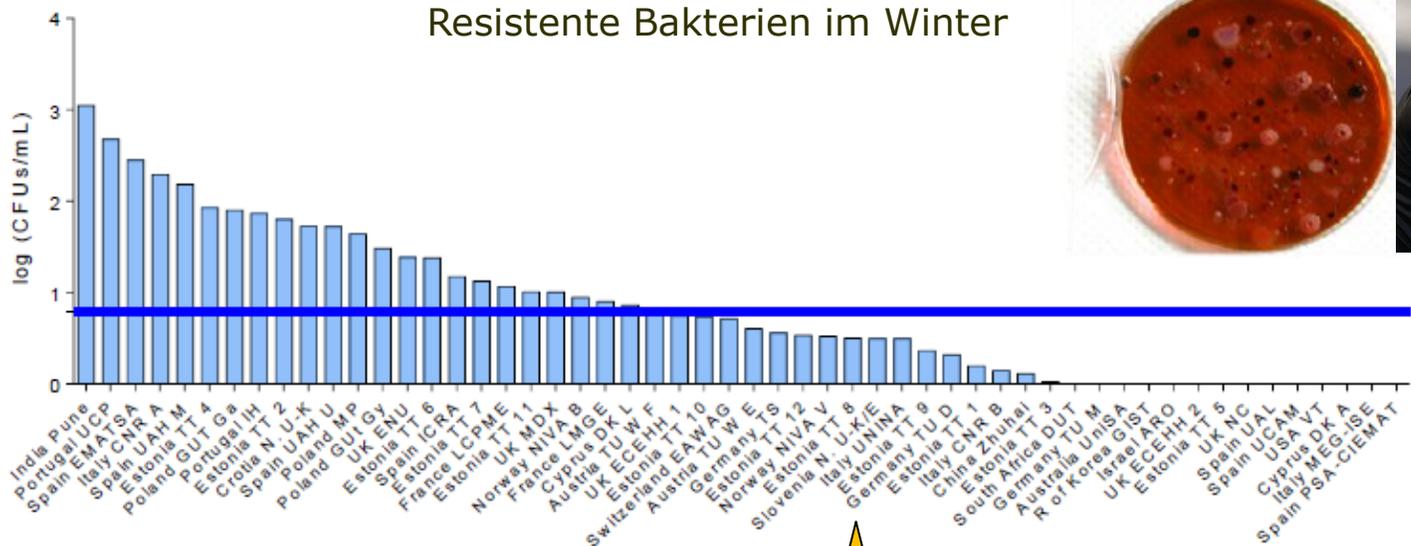


resistente Bakterien
behandelbare Bakterien

Norhan et al. Scientific Reports 2018

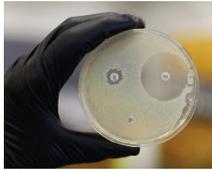
Mögliche Methoden des AMR monitoring

Kontrolle - Monitoring



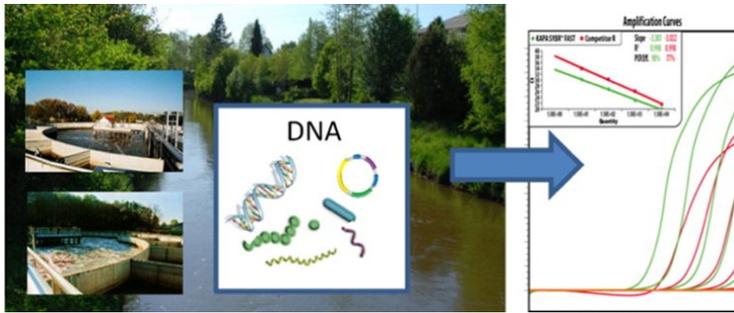
Marano et al. 2020 Environment International

Mögliche Methoden des AMR monitoring



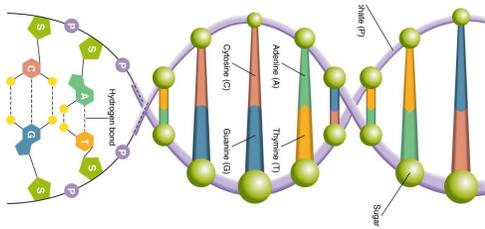
- Kulturbasiert (langsam) ✓
- Molekular

⇒ Quantifizierungsanalysen (schnell)



- Quantifikation Resistenzgene
- Q-PCR – high throughput qPCR
- digitale PCR

- Sequenzierungstechnologien (hoher bioinformatischer Aufwand)
- Metagenomik (long or short read)
- Epic-PCR
- Hi-C linkage



- Sensor Technologien (erst am Anfang)

⇒ Optical, DNA or Proteinbased

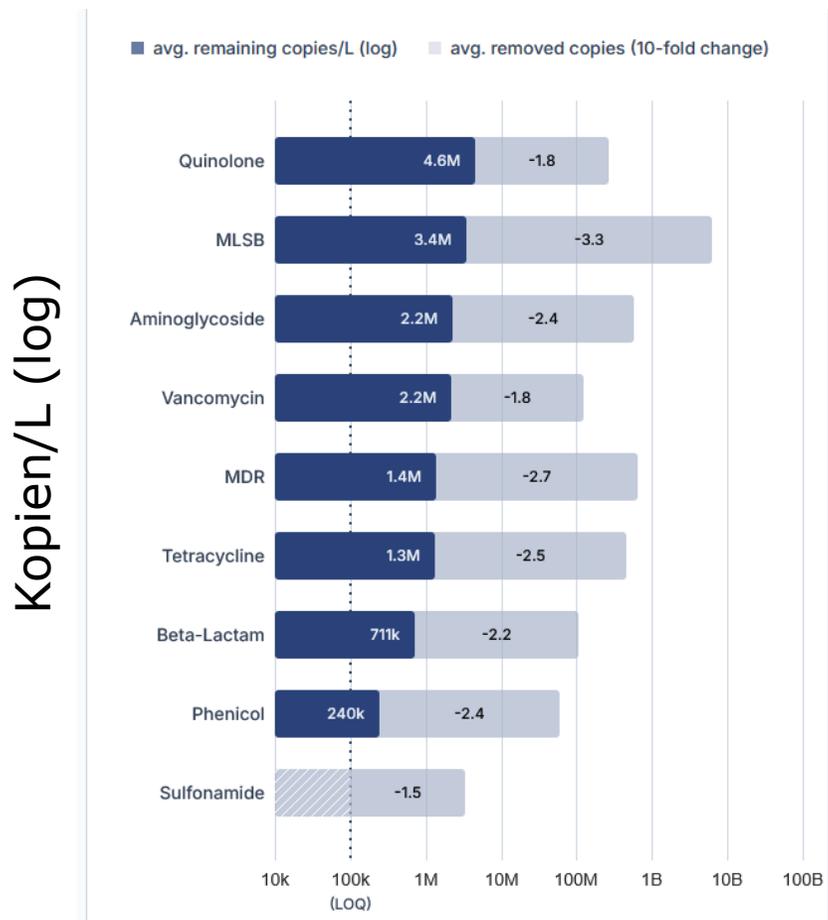


Nochmal:

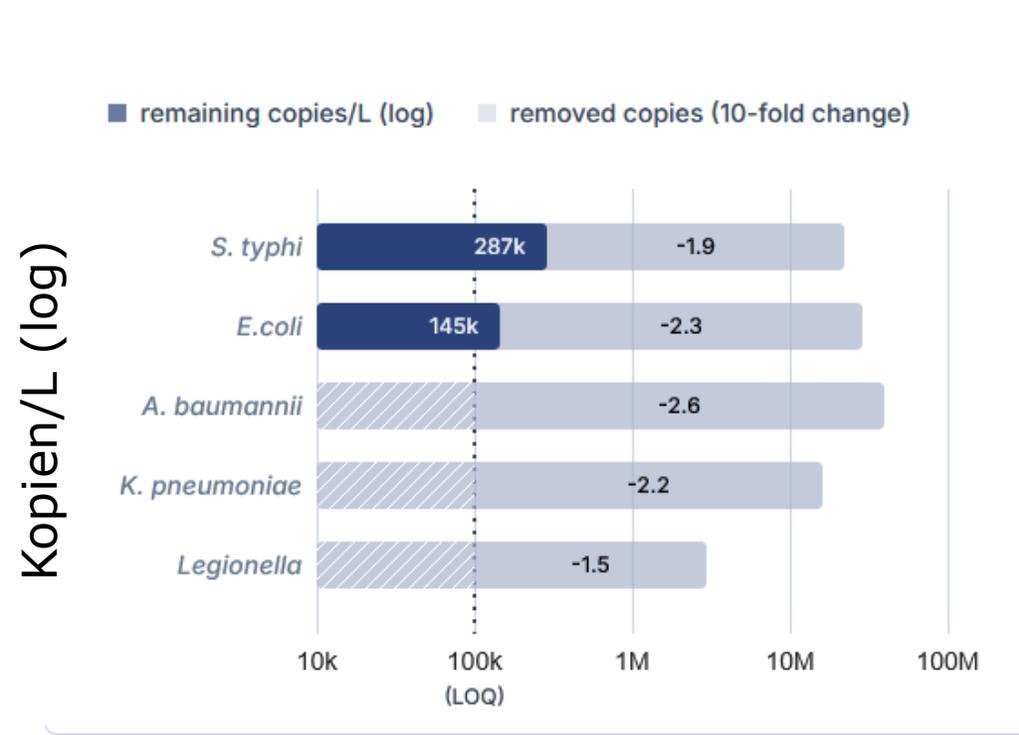


Quantifizierung der Resistenzgene und der Pathogene

Reduktion der Resistenzgene



Pathogene Bakterien



Zeitplan

ChatGPT

Die Richtlinie trat am 1. Januar 2025 in Kraft.
Die Mitgliedstaaten müssen die Bestimmungen bis spätestens 31. Juli 2027 in nationales Recht umsetzen.

Die Kommission erlässt bis zum **2. Juli 2026 Durchführungsrechtsakte**, um eine Mindesthäufigkeit für Probenahmen und eine harmonisierte Methode zur Messung antimikrobieller Resistenzen im kommunalen Abwasser festzulegen, wobei sie mindestens alle verfügbaren Daten der nationalen Gesundheitsbehörden und der für die Überwachung antimikrobieller Resistenzen zuständigen nationalen Behörden berücksichtigt. Diese Durchführungsrechtsakte werden gemäß dem in Artikel 28 Absatz 2 genannten Prüfverfahren erlassen.

Artikel 22

Informationen über die Überwachung der Durchführung

- a) bis zum **31. Dezember 2028 einen Datensatz mit Informationen, die gemäß Artikel 21** erhoben wurden, einschließlich Informationen über die Parameter gemäß Artikel 21 Absatz 1 Buchstabe a und die Prüfergebnisse in Bezug auf die in Anhang I Teil C festgelegten Kriterien für die Erfüllung/Nichterfüllung der Anforderungen, und aktualisieren diesen Datensatz danach jährlich, ???
- h) bis zum **31. Dezember 2030 einen Datensatz mit den in Artikel 17** Absätze 1 und 3 genannten Überwachungsergebnissen und aktualisieren diesen Datensatz danach jährlich,



Zusammenfassung - Ausblick

- Das Monitoring antimikrobieller Resistenzen wird auf kulturbasierten und molekularen Methoden basieren.
- Eine Standardisierung der Methoden ist unerlässlich
- (dringender) Bedarf an einem einheitlichen AMR-Datenmanagementsystem für den Umweltbereich, das eine sektorübergreifende Datenintegration ermöglicht und Forscher, Aufsichtsbehörden und Betreiber von Kläranlagen unterstützt.

My group



GEFÖRDERT VOM



Bundesministerium
für Bildung
und Forschung

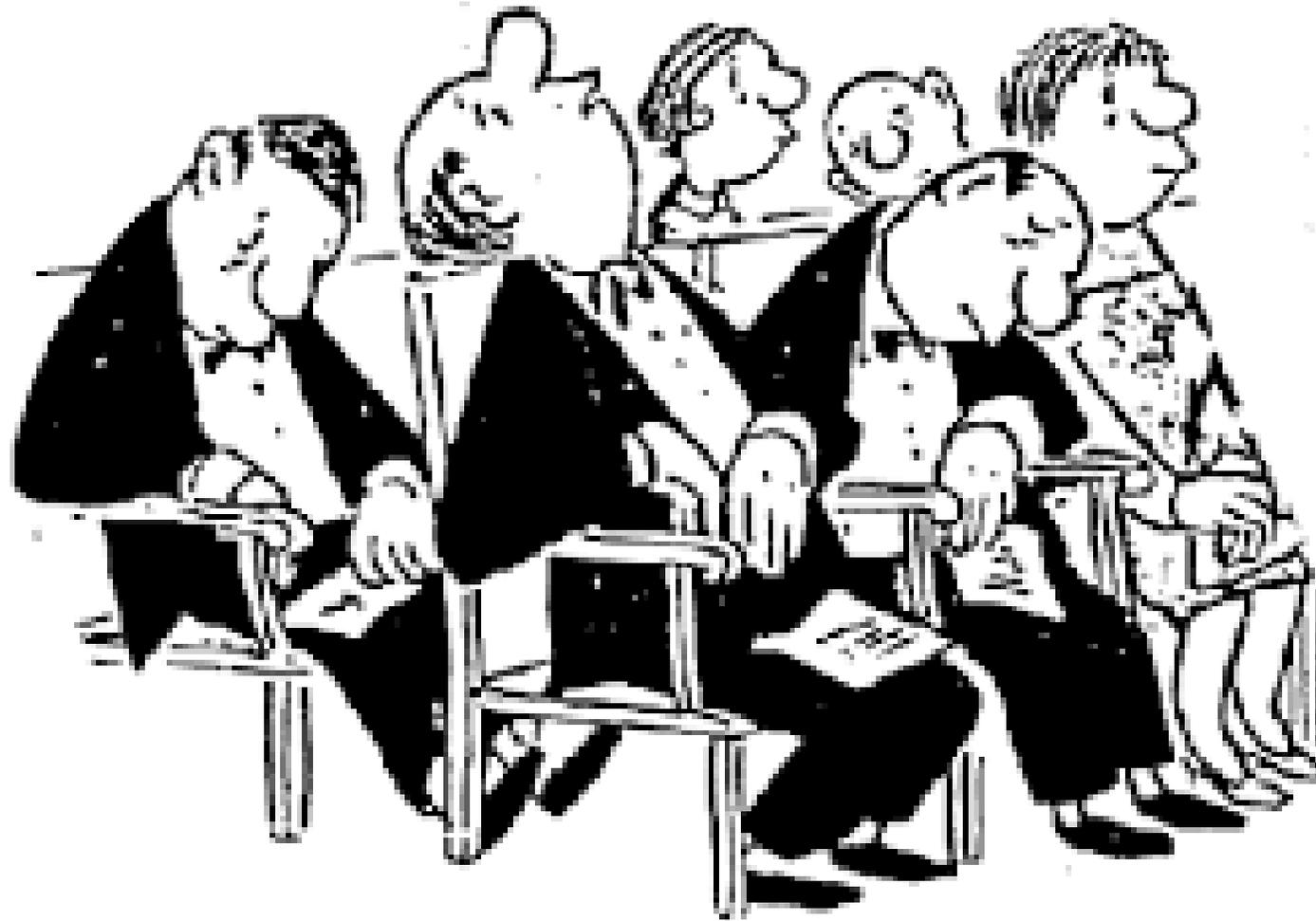


01KI2401 EXPLORE
01KI2128 ACRAS-R

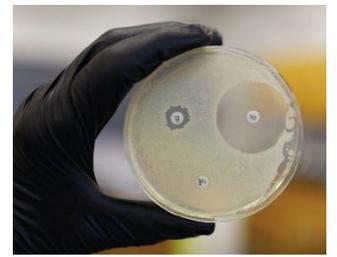
JPI AMR EMBARK
JPI AMR – TEXAS, SEARCHER



Danke für die Aufmerksamkeit

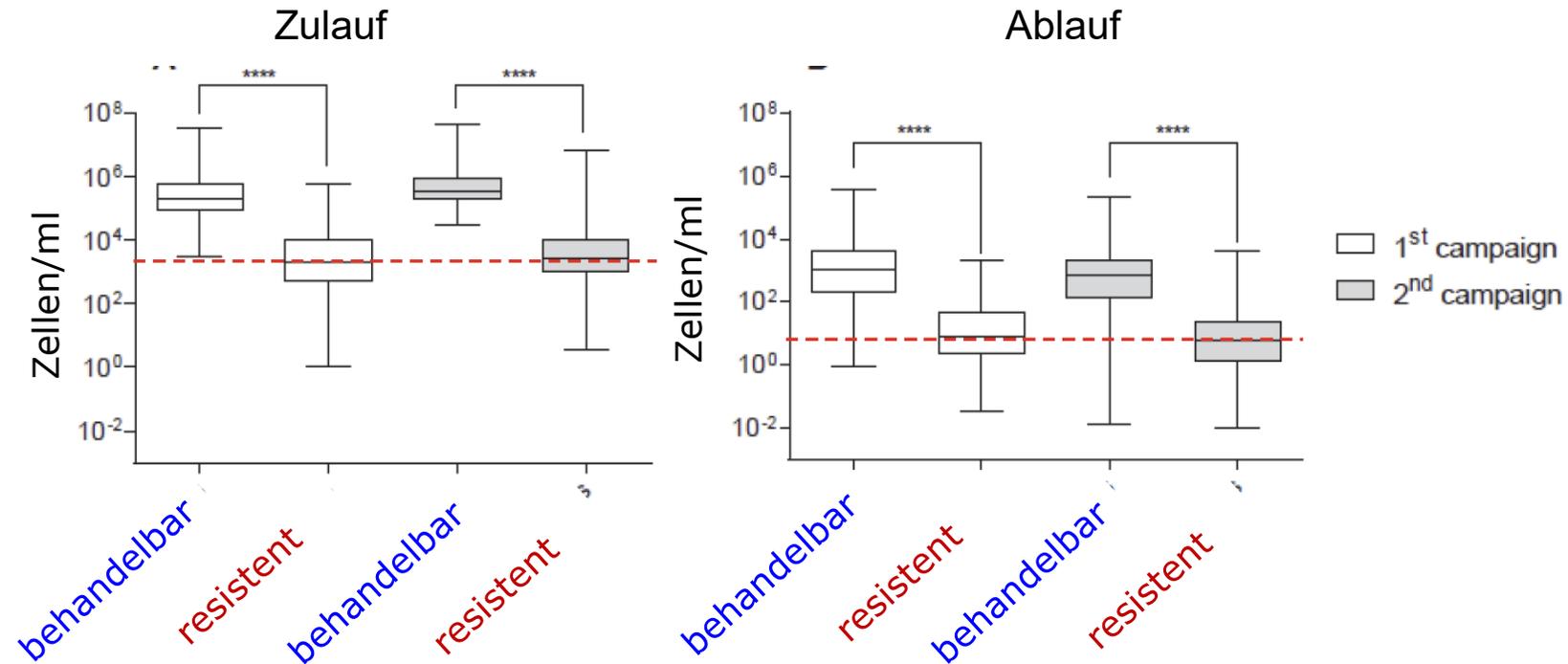


Mögliche Methoden des AMR monitoring



- Kulturbasiert

⇒ Cefotaxim Resistente Bakterien (Coliforme) – WWTP globale Studie (54 WWTP)



Marano et al. 2020 Environment International



The ISME Journal, 2024, 18(1), wrae243

<https://doi.org/10.1093/ismejo/wrae243>

Advance access publication: 10 December 2024

Original Article

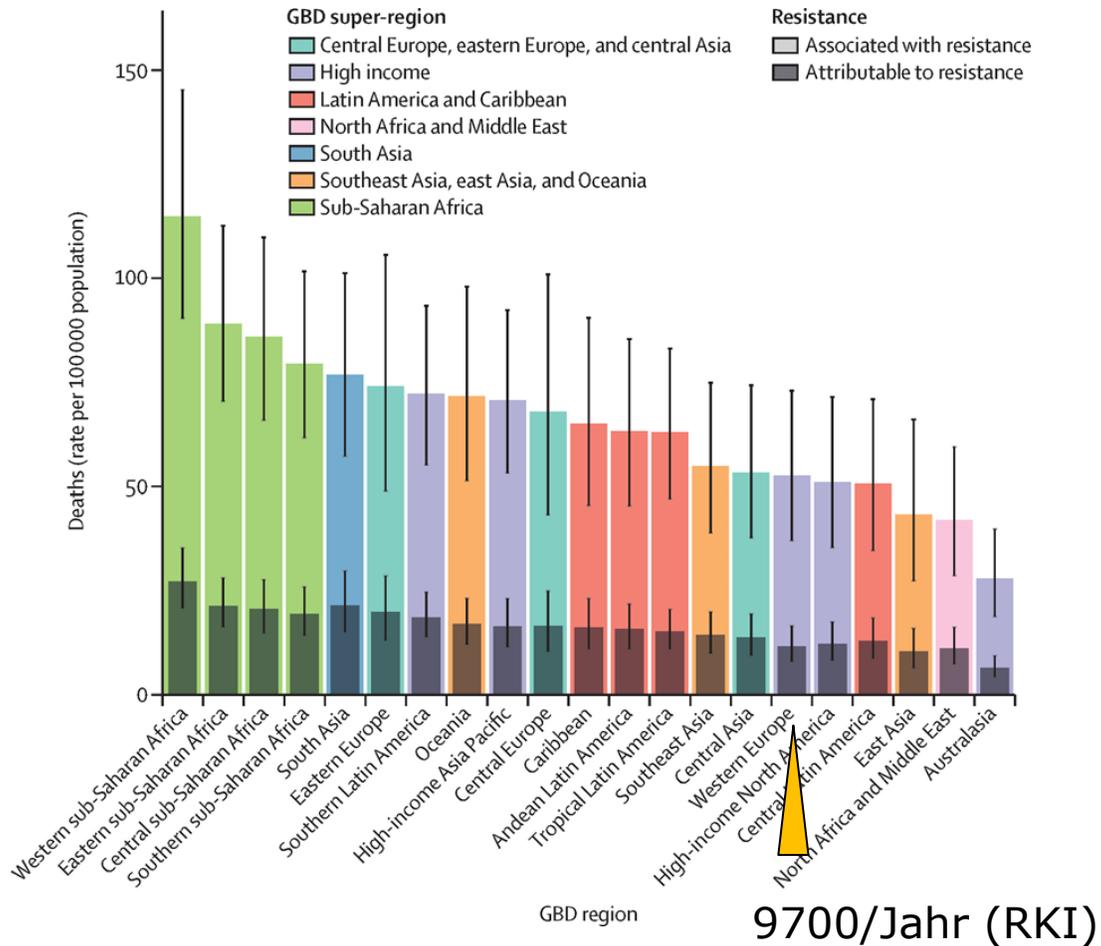
Bacteria of the order *Burkholderiales* are original environmental hosts of type II trimethoprim resistance genes (*dfrB*)

David Kneis*, Faina Tskhay, Magali de la Cruz Barron, Thomas U. Berendonk

Dresden University of Technology, Institute of Hydrobiology, 01062 Dresden, Saxony, Germany

<https://doi.org/10.1093/ismejo/wrae243>

„Die Stille Pandemie“



The Lancet 2022 399629-655DOI: (10.1016/S0140-6736(21)02724-0)

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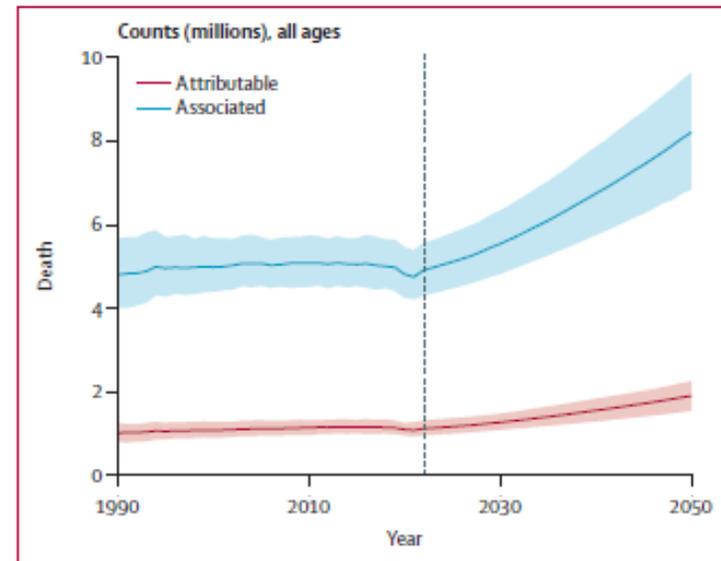


Figure 6: Global attributable and associated AMR burden in the reference scenario, 2022–2050

Shading represents the 95% uncertainty interval. The vertical line is placed at 2021 to distinguish estimates from forecasts.

Global burden of bacterial antimicrobial resistance 1990–2021: a systematic analysis with forecasts to 2050

GBD 2021 Antimicrobial Resistance Collaborators*

www.thelancet.com Published online September 16, 2024 [https://doi.org/10.1016/S0140-6736\(24\)01867-1](https://doi.org/10.1016/S0140-6736(24)01867-1)

Mögliche Methoden des AMR monitoring

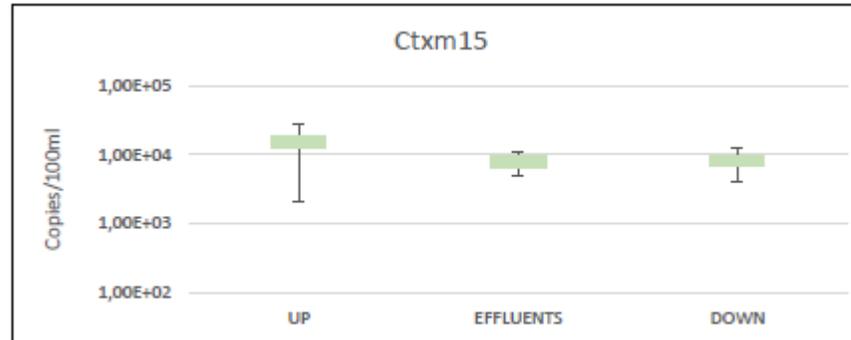
- Kulturbasiert
- Molekular
 - ⇒ Quantifikation Resistenzgene
 - ⇒ Q-PCR – high throughput qPCR
 - ⇒ digitale PCR

 - ⇒ Sequenzierungstechnologien (+Einzellzellsequenzierung)
 - ⇒ Metagenomik (long or short read)
 - ⇒ Epic-PCR
 - ⇒ Hi-C linkage
- Sensor Technologien
 - ⇒ Optical, DNA or Proteinbased

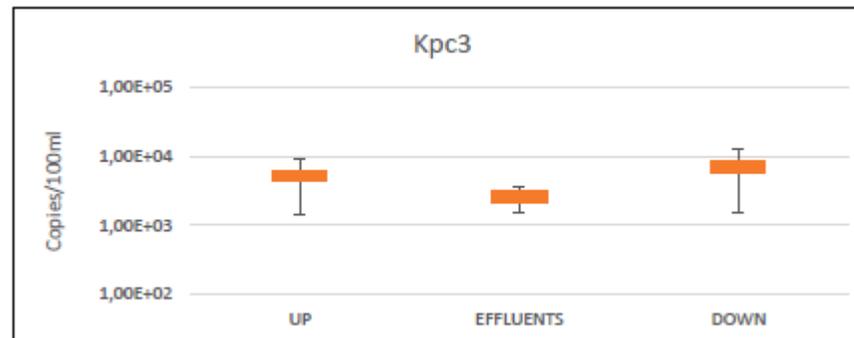
Surveillance of resistance in the environment

The receiving water

β -Lactamase - Penicillin



β -Lactamase – Penicillin *K. pneumoniae*



Mögliche Methoden des AMR monitoring

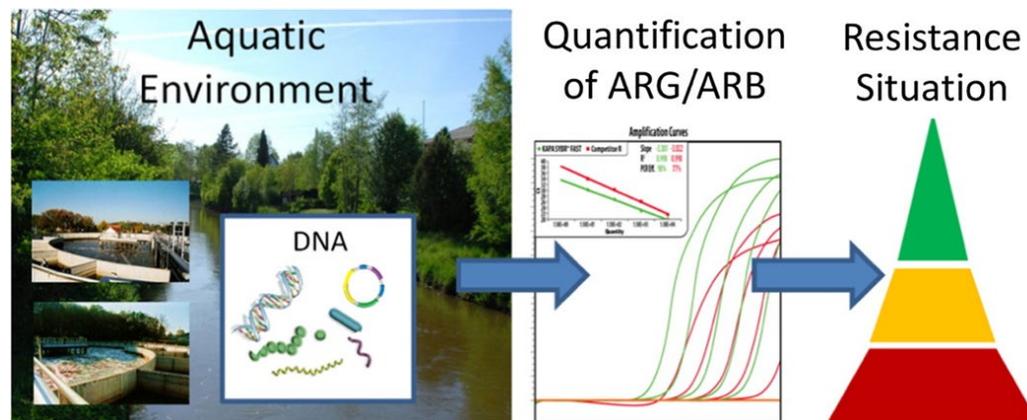
- Kulturbasiert

⇒ Bsp.: *Escherichia coli*

- DIN EN ISO 9308-2:2014 DE MPN
- Ursprüngliche Motivation eine Fäkalverunreinigung zu detektieren
- Geeignet um eine *E.coli* – Coliforme Belastung abzuschätzen ?
- Resistenten Anteil abschätzen ?

Mögliche Methoden des AMR monitoring

- Molekular
 - ⇒ Quantifikation Resistenzgene
- Quantifizierung als Anzahl der Genkopien pro 16S („Bakterienmasse“)
 - ⇒ „relative Anzahl der Resistenzgene“
- Quantifizierung als Anzahl der Genkopien per Volumen
 - ⇒ „Absolute Anzahl der Resistenzgene“ (Konzentration)



Rocha et al. 2018 J. Environ. Chem. Eng



Article

<https://doi.org/10.1038/s41467-022-34312-7>

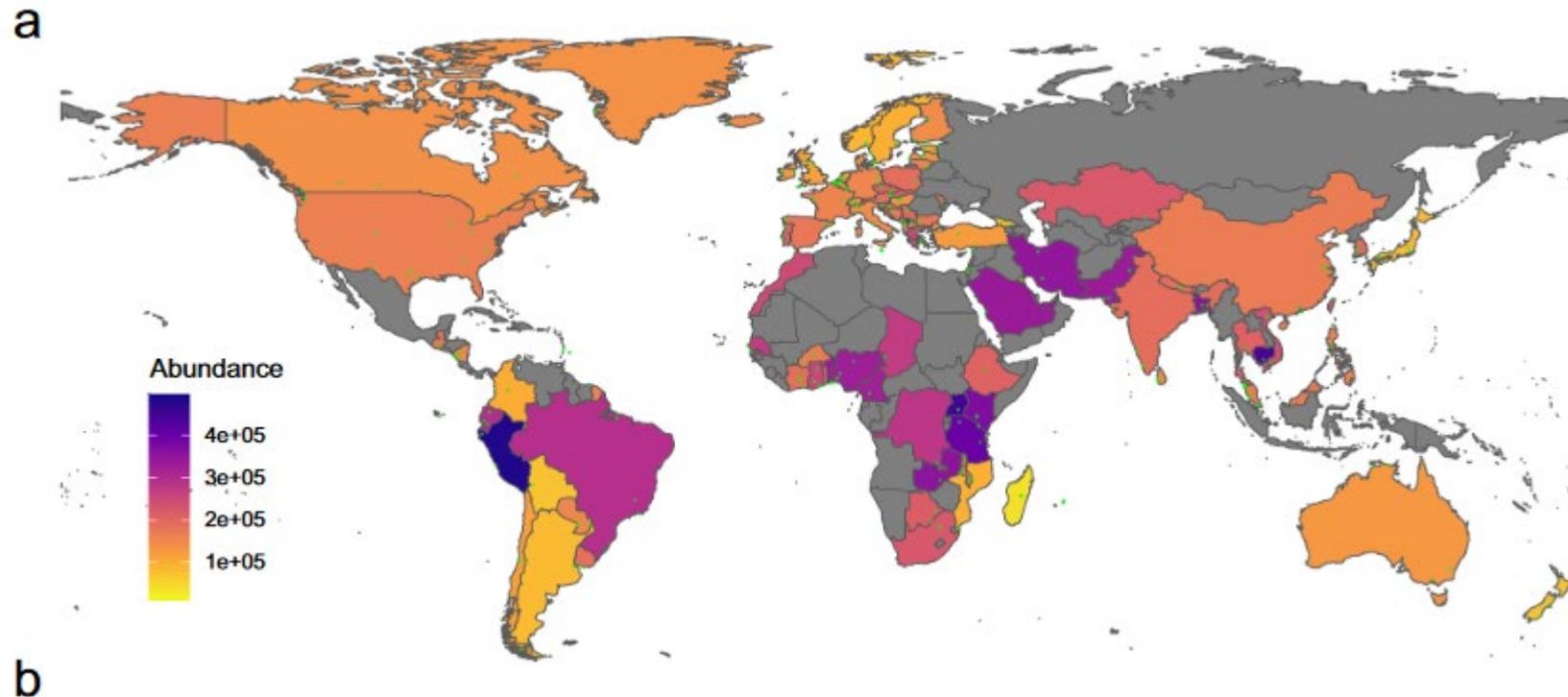
Genomic analysis of sewage from 101 countries reveals global landscape of antimicrobial resistance

Received: 25 July 2022

Accepted: 20 October 2022

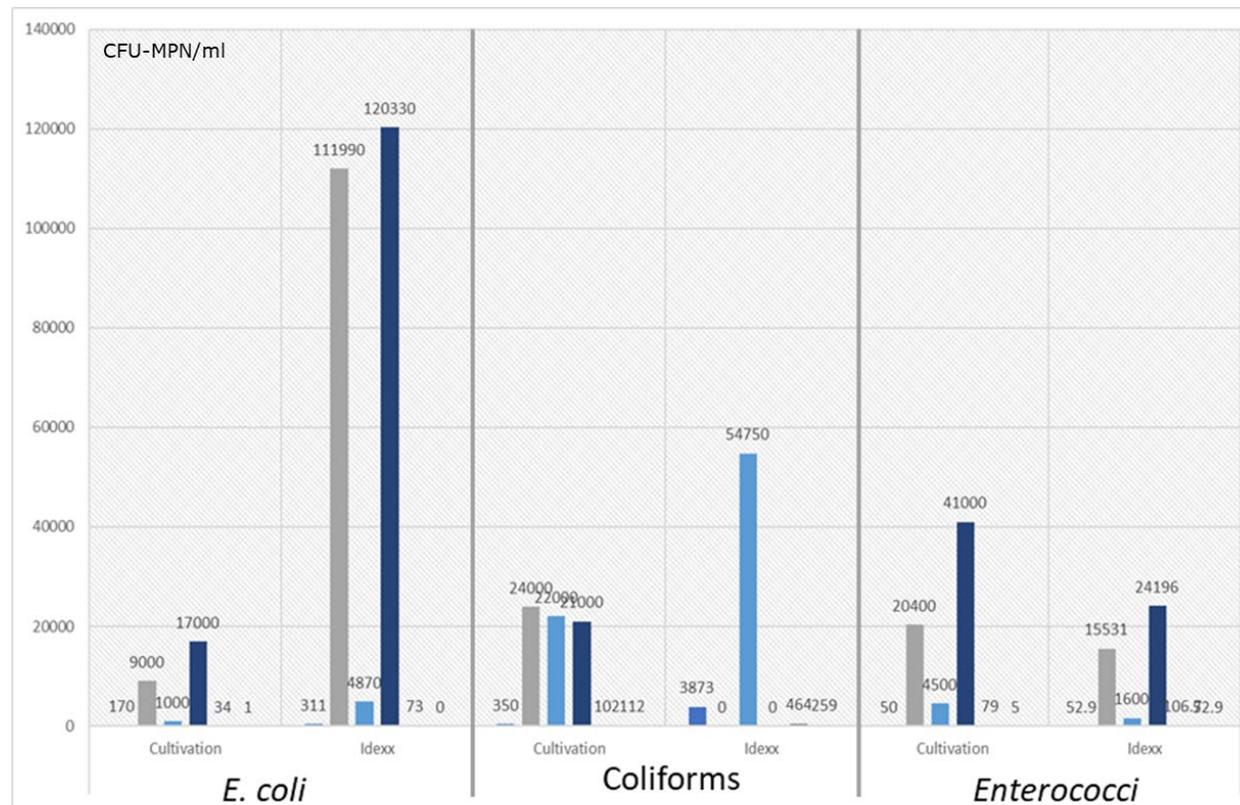
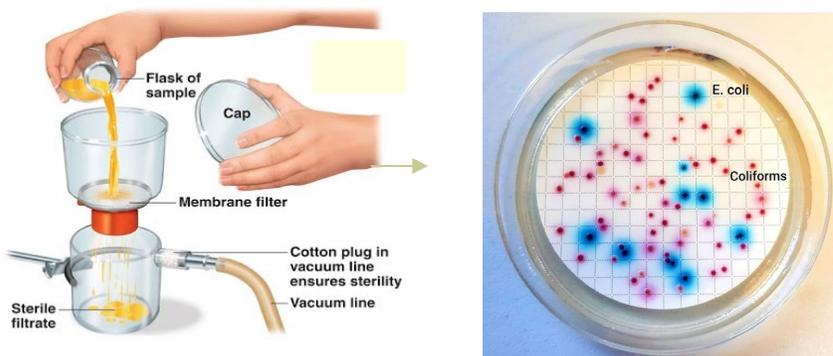
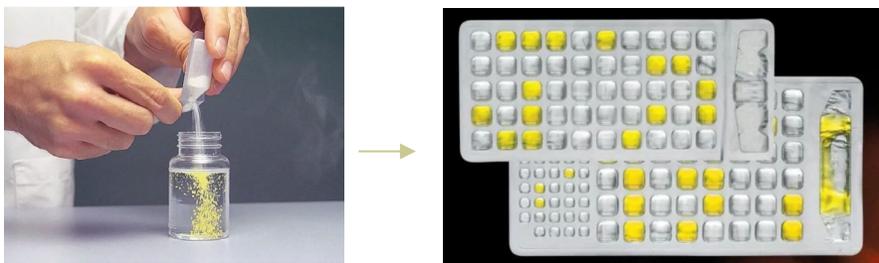
Published online: 01 December 2022

Patrick Munk¹✉, Christian Brinch¹, Frederik Duus Møller¹,
Thomas N. Petersen¹, Rene S. Hendriksen¹, Anne Mette Seyfarth¹,
Jette S. Kjeldgaard¹, Christina Aaby Svendsen¹, Bram van Bunnik²,
Fanny Berglund³, Global Sewage Surveillance Consortium*



- Kulturbasiert Mögliche Methoden des AMR monitoring

- ⇒ Escherichia coli – coliforme
- ⇒ Vergleich Chomagar - Colilert

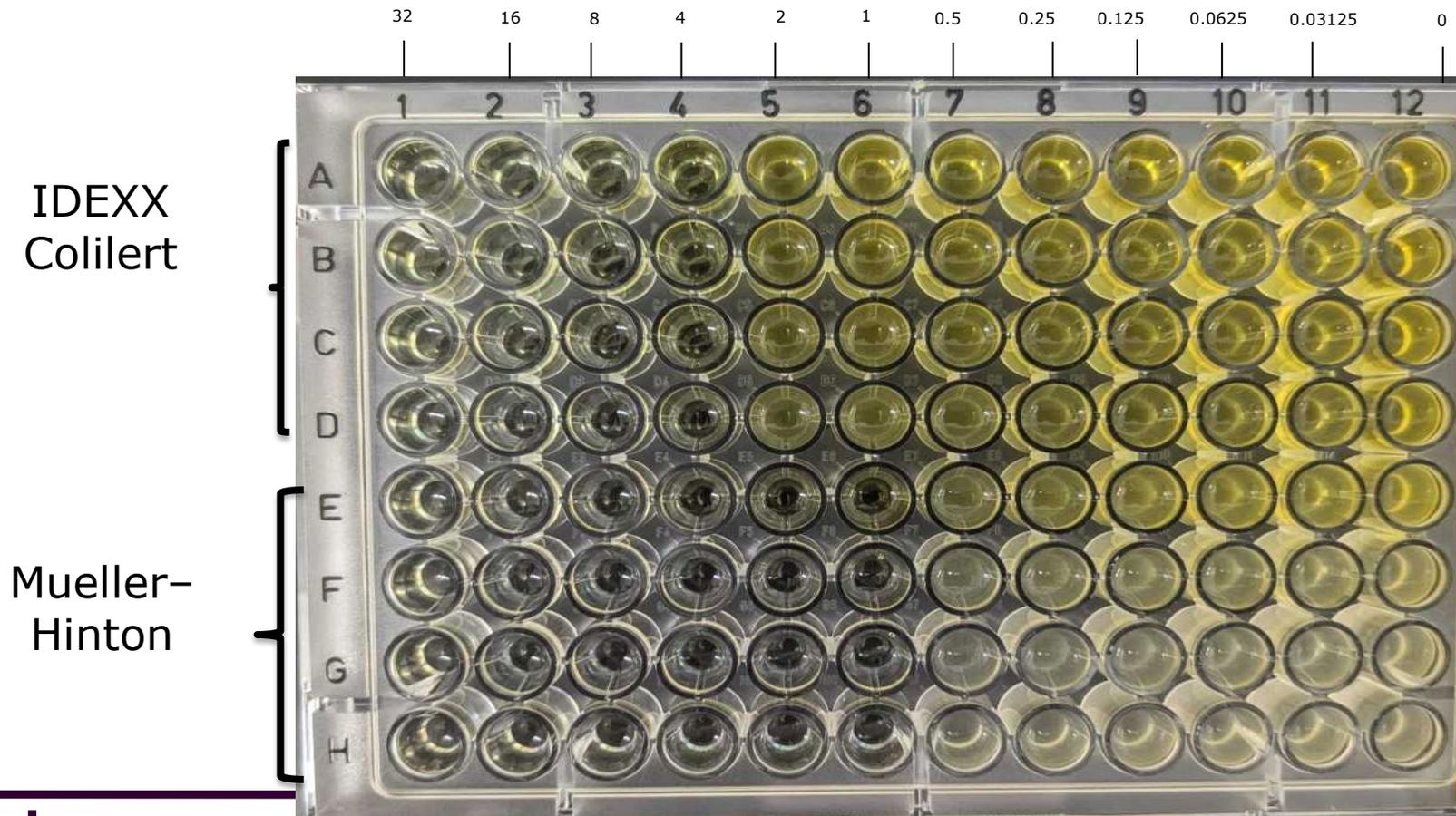


Unterschiedliche Farben sind verschiedene Probenahmestellen an der Kläranlage

Mögliche Methoden des AMR monitoring

- Kulturbasiert

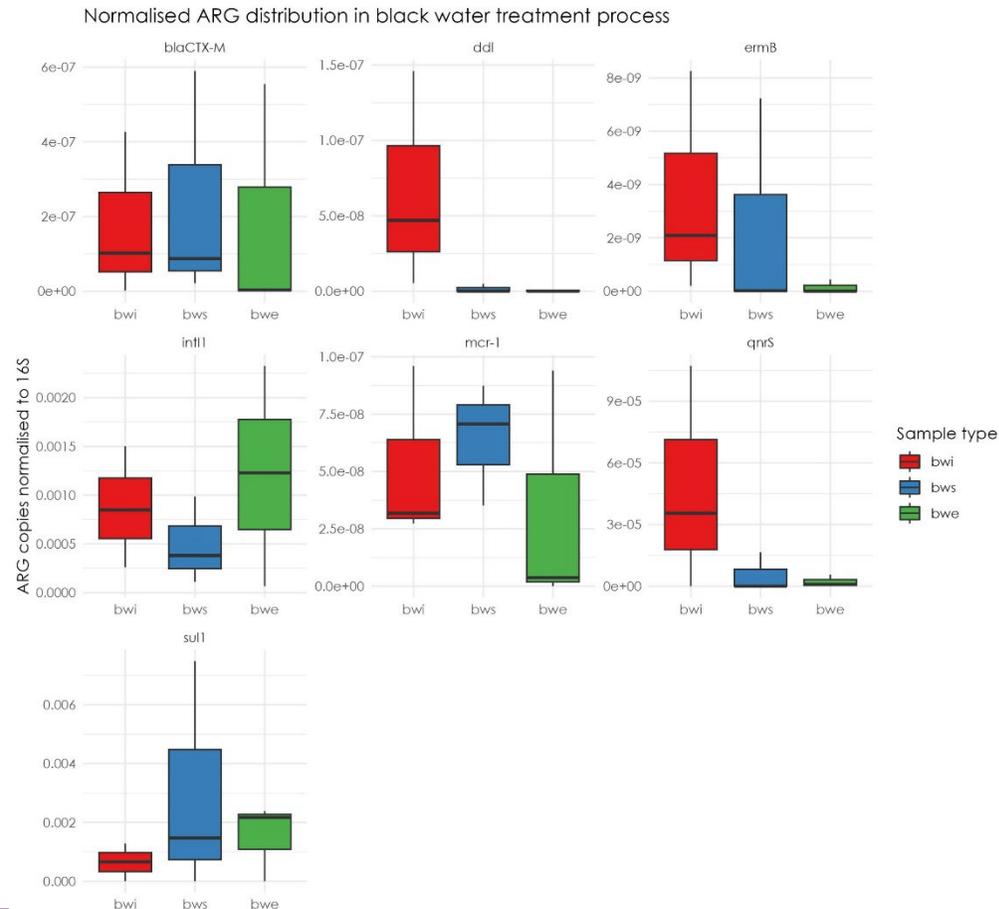
⇒ Anpassung des Colilert an AMR Detektion schlug fehl



Mögliche Methoden des AMR monitoring

- Molekular
 - ⇒ Quantifikation Resistenzgene

- Beispiel: Anaerobic MBR Spanien



BW	
gene	Removal (%)
<i>bla_{CTX-M}</i>	50
<i>ddl</i>	~100
<i>ermB</i>	62
<i>int11</i>	Increase
<i>mcr-1</i>	37
<i>qnrS</i>	95
<i>sul1</i>	Increase

relevante Parameter für die öffentliche Gesundheit zu ermitteln, die zumindest im Zulauf kommunaler Abwasserbehandlungsanlagen zu überwachen sind, unter Berücksichtigung der verfügbaren Empfehlungen unter anderem des Europäischen Zentrums für die Prävention und die Kontrolle von Krankheiten (ECDC), der Europäischen Behörde für die Krisenvorsorge und -reaktion bei gesundheitlichen Notlagen (HERA) und der Weltgesundheitsorganisation (WHO),

Mögliche Methoden des AMR monitoring

Kontrollen - Behandlungstechnologien

DSWAP Removal Efficiency Database

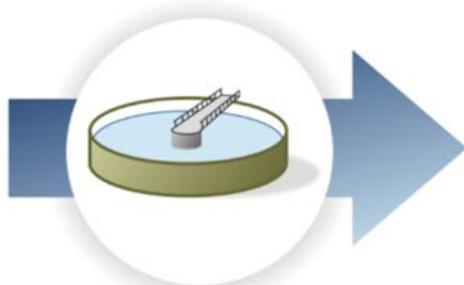
Start page | High-level summaries | Tabular views | SQL mode | Impressum | Legal aspects

This is the web interface to the database of removal efficiencies observed in wastewater treatment. It represents the knowledge reported in the scientific literature as of 2022 with regard to selected chemical or biological contaminants and treatment technologies.

The unit of values reported differs between chemical contaminants (e.g. antibiotics or other drugs) and biological items (such as genetic markers). The values are computed from concentrations (or loads) in a treatment module's input (C_{in}) and output (C_{out}) as follows:

Contaminant group	Reported value	Unit	Definition
Chemical	Percentage removal efficiency	%	$PRE = (C_{in} - C_{out}) / C_{in} * 100$
Biological	Log removal value	-	$LRV = \log_{10}(C_{in} / C_{out})$

Use the navigation tabs at the top of the page to check the different functions of this web interface. For security reasons, the provided features differ between an online installation of this app (server mode) and local installations.



Mögliche Methoden des AMR monitoring

Kontrollen - Behandlungstechnologien

DSWAP Removal Efficiency Database

Start page | High-level summaries | Tabular views | SQL mode | Impressum | Legal aspects

Numbers of records | Ranges per contaminant | Ranges per technology group

Select criterion for building the summary

TechnologyGroup

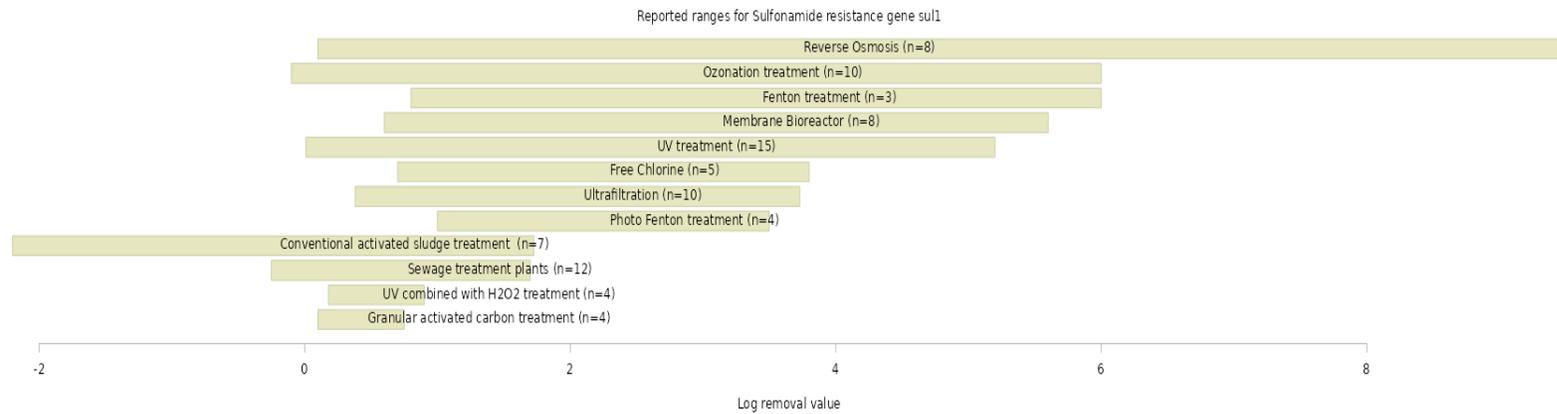
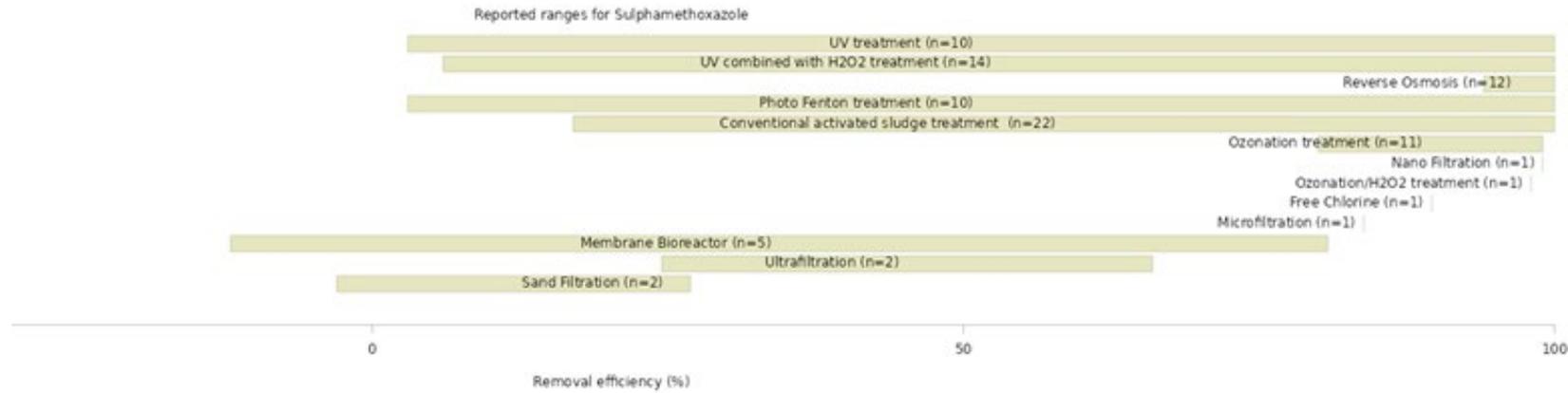
TechnologyGroup	Records
Conventional activated sludge treatment	114
Fenton treatment	5
Forward Osmosis	3
Free Chlorine	25
Granular activated carbon treatment	22
Membrane Bioreactor	46
Microfiltration	6
Nano Filtration	7
Ozonation treatment	68
Ozonation/H2O2 treatment	4
Photo Fenton treatment	47
Reverse Osmosis	36
Sand Filtration	15
Sewage treatment plants	36
UV combined with H2O2 treatment	94

SCAN ME

https://limno-live.hydro.tu-dresden.de/dkneis/prototypes/dswap/#tab-4270-2

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Zusammenfassung und Schlussfolgerungen

- Antibiotikaresistenz im Abwasser ist ein relevantes Problem
- Zum Monitoring bestimmter Kontrollmaßnahmen (e.g. Aktivkohle, Membran, Ozon) stehen unterschiedliche Werkzeuge zur Verfügung
 - ⇒ Datenlage zu Maßnahmen ist noch unübersichtlich
- Kulturbasierte Ansätze (Leitl. EU-Verordnung 2020/741, EU WasserWVVO 2022/C 2989/01)
 - ⇒ entsprechende Indikatorsysteme existieren (ABER müssen evtl. angepasst werden)
 - ⇒ Wirt – Resistenzgen Verhältnis am besten dargestellt
 - ⇒ Im klinischen Bereich existieren Normierungen
- Q-PCR (real time, chip, dd-PCR) Berendonk et al. 2015, Abramova et al. 2023 Alygizakis et al. 2024
 - ⇒ Analytisch schnellster Ansatz
 - ⇒ gut zu normieren (e.g. ISO)
 - ⇒ Beste Sensitivität für "niedrige" Konzentrationen
- Metagenomik

Monitoring – (passive ?) Surveillance

Was nun ?

- Derzeit am wahrscheinlichsten das sich entweder ein kulturbasierter oder ein mit q-PCR kombinierter Ansatz durchsetzt.
- In den USA scheint die EPA aufgrund der Covid-Analysen eine q-PCR System zu etablieren.

DANKE an meine AG, besonders Dr. Klümper, Dr. Kneis, Dr. De LaCruz, Mrs. Faina Tskhay



NORMAN-network

JPI-Water - PRESAGE
JPI AMR EMBARK, SEARCHER
JPI Biodiversa - ANTIVERSA

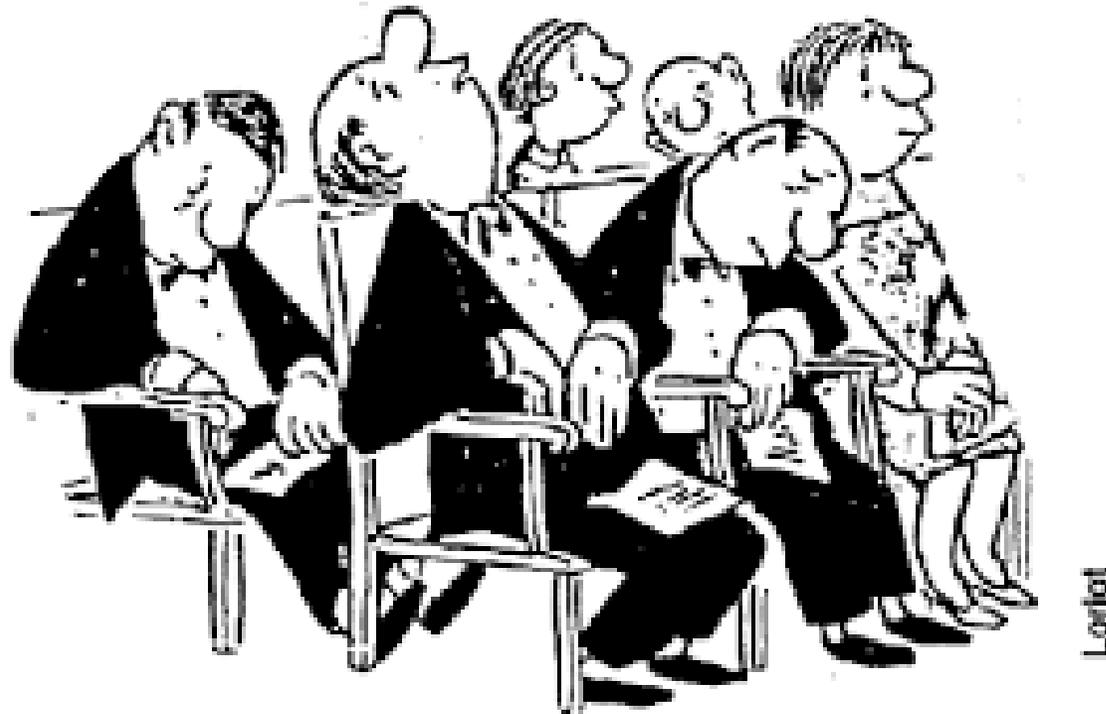


GEFÖRDERT VOM

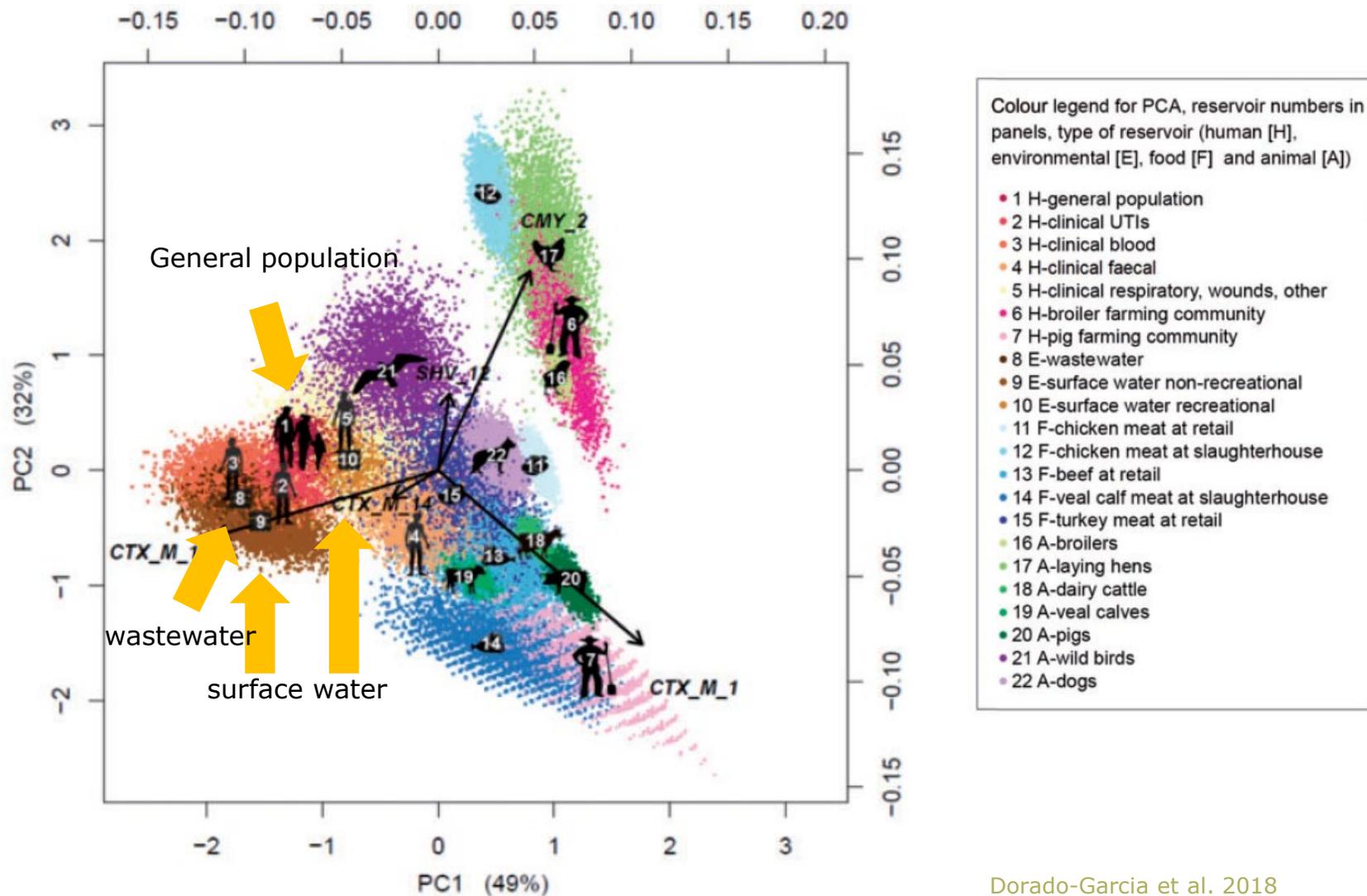
Bundesministerium
für Bildung
und Forschung



Danke für Ihre Aufmerksamkeit



Resistant Escherichia coli and their genome association



Dorado-Garcia et al. 2018

Was bedeuten 10^{-3} resistance gene /16S/ml ?
 Jedes 1000 te Bacterium besitzt ein Resistenzgen (ARG)

Bakterienkonzentration im Ablauf : $\sim 10^6$ Zellen/ml
 -> 10^3 Zellen mit einem Resistenzgen/ml ($10^6/L$)

Volumen des Ablaufs: $\sim 10^8$ L/d (10^5 m³)
 -> 10^{14} Zellen mit einem Resistenzgen /d

Bacterium mit ARG : Umweltbakterium
 1 : 10 - 10000

oder

10^4 Resistenzgene (ARG) pro ml im Kläranlagenablauf ?

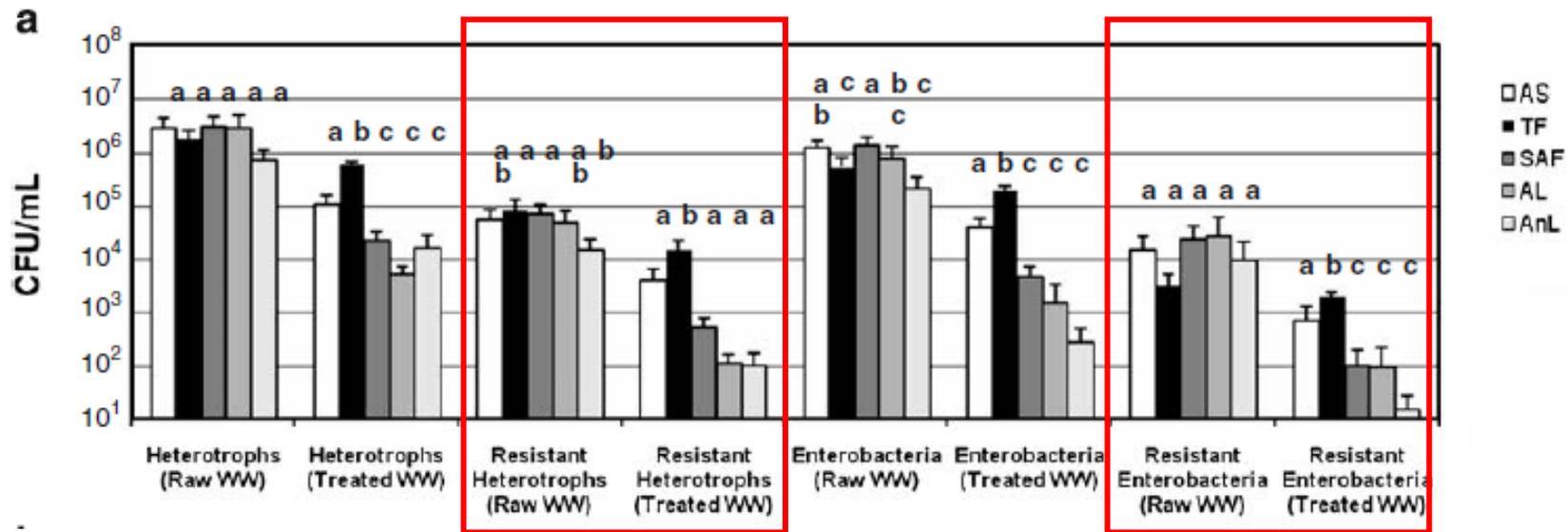
c. 10^{15} Resistenzgenkopien (pro Gentyp) pro Tag verlassen die Kläranlage

Ciprofloxacin Resistance in Domestic Wastewater Treatment Plants

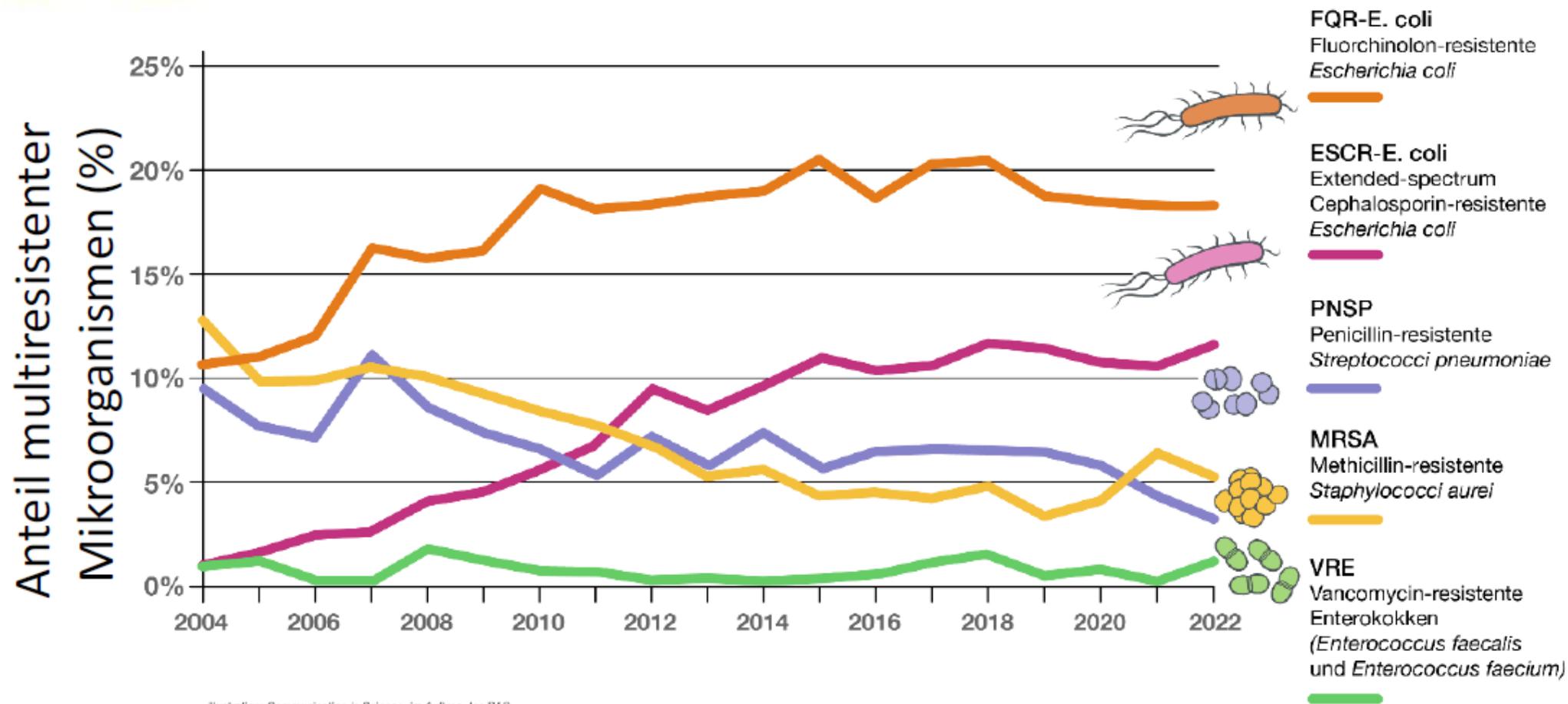
Celia M. Manaia • Ana Novo • Bruno Coelho • Olga C. Nunes

Water Air Soil Pollut (2010) 208:335–343

339



„ Independently of the size or type of treatment used domestic wastewater treatment plants discharged per day at least 10^{10} – 10^{14} colony forming units of ciprofloxacin-resistant bacteria“



B. Surveillance and monitoring of AMR and antimicrobial consumption (AMC)

HEREBY ENCOURAGES MEMBER STATES TO:

5. Close existing surveillance and monitoring gaps and ensure completeness of data, including real-time data and timely access to data where appropriate by 2030, on both AMR and AMC at all levels (e.g. community, hospitals and long-term care facilities) to support the prudent use of antimicrobials in human health, by:
 - a. ensuring, in coordination with ECDC, that surveillance of AMR in bacteria from humans encompasses not only bloodstream and cerebrospinal fluid isolates (invasive isolates) but also all other isolates from clinical microbiology laboratories, and that the corresponding data are regularly reported to the ECDC to rapidly detect and better gauge the scale and spread of antimicrobial resistant pathogens within and across Member States;
 - b. requiring, taking into account any methodology established at EU level, that infections caused by critical (high negative health impact) multidrug-resistant organisms resistant to last line treatments, e.g. carbapenem-resistant *Acinetobacter baumannii*, carbapenem-resistant Enterobacteriaceae (e.g. *Klebsiella pneumoniae*, *Escherichia coli*) and *Candida auris*, are notifiable diseases under national legislation. Member States can decide if other resistant organisms are notifiable, according to the national situation and need;
 - c. expanding surveillance of AMR in humans to pathogens with emerging or established AMR, due to their exposure to substances in the environment, in particular those used in plant protection products or biocidal products;
 - d. collecting data on AMC, in humans at the appropriate levels to allow the monitoring of antimicrobial prescribing and to provide timely feedback on prescription trends and patterns involving, among others, prescribers, pharmacists and other parties collecting such data, and where possible and appropriate using EU level digital infrastructure.
 - e. developing integrated systems for the surveillance of AMR and AMC encompassing human health, animal health, plant health, food, wastewater and the environment (in particular water and soil), taking into account the Commission feasibility study on integrated systems, the work of the Quadripartite QTS-AIS expert group on Integrated Surveillance⁽⁵⁶⁾ as well as other initiatives already launched, such as the WHO Tricycle protocol for an integrated global surveillance on ESBL-producing *E. coli* across the human, animal and environmental sectors. Such integrated and continuous intersectoral monitoring should be designed to efficiently and rapidly detect emerging resistant infections and outbreaks but equally as regards soil and water bodies to determine the presence of AMR genes and antimicrobials, the trends and their toxicity. The results of this surveillance should inform effective strategies to tackle AMR across sectors and at appropriate administrative levels.

Commission Notice Guidelines to support the application of Regulation 2020/741 on minimum requirements for water reuse 2022/C 298/01
C/2022/5489

OJ C 298, 5.8.2022, p. 1–55 (BG, ES, CS, DA, DE, ET, EL, EN, FR, GA, HR, IT, LV, LT, HU, MT, NL, PL, PT, RO, SK, SL, FI, SV)

6. **Additional requirements (KRM6)** - the outcomes of the risk assessment might identify additional or stricter water quality and monitoring requirements than those from Section 2 of Annex I of the Regulation.

If additional parameters or limits are included, this should be based on the outcomes of the risk assessment and supported by scientific evidence that they originate from the water reuse system and not from other sources.

These additional parameters may also include the following pollutants: heavy metals, pesticides, disinfection by-products, pharmaceuticals, substances of emerging concern, bacteria that exhibit anti-microbial resistance.

(23) The Union recognises the importance of tackling the issue of antimicrobial resistance (AMR) and adopted in 2017 the European One Health Action Plan against AMR⁵⁵. According to the World Health Organisation (WHO), wastewater is recognised and documented as major sources of antimicrobial agents and their metabolites, as well as antimicrobial-resistant bacteria and their genes. In order to increase the knowledge on the main sources of AMR, it is necessary to introduce a monitoring obligation for the presence of AMR in urban wastewaters to further develop our scientific knowledge and potentially take adequate action in the future.

Proposal for a

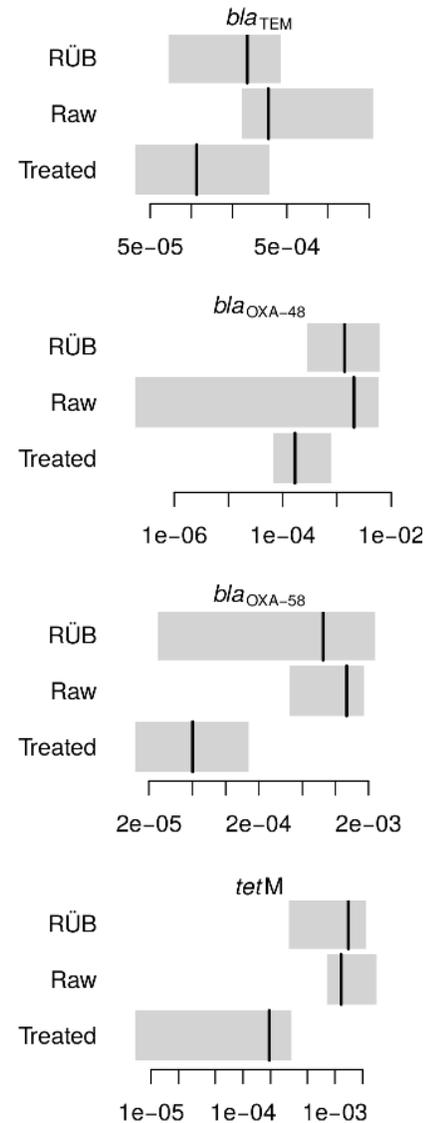
DIRECTIVE OF THE EUROPEAN PARLIAMENT AND OF THE COUNCIL

concerning urban wastewater treatment (recast)

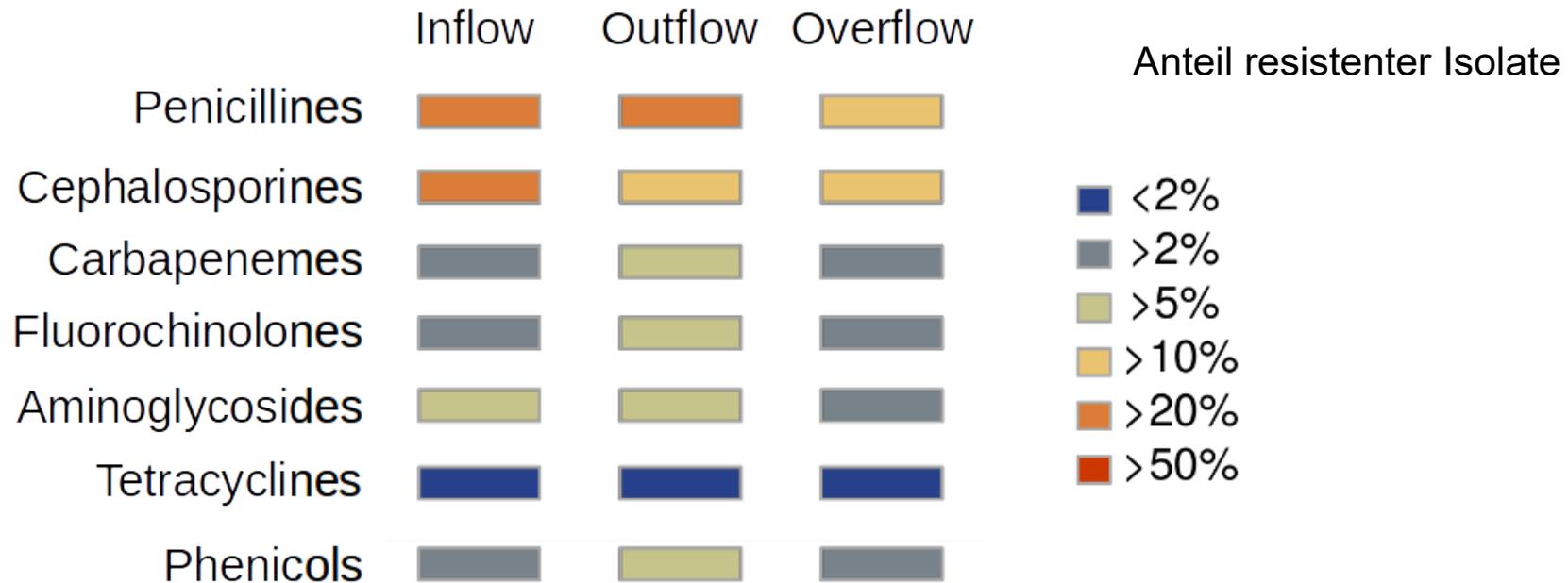
4. For agglomerations of 100 000 p.e. and above, Member States shall, by 1 January 2025, ensure that antimicrobial resistance is monitored at least twice a year at the inlets and outlets of urban wastewater treatment plants and, when relevant, in the collecting systems.

The Commission shall adopt implementing acts in accordance with the procedure referred to in Article 28 to ensure an uniform application of this Directive by establishing a harmonised methodology for measuring antimicrobial resistance in urban wastewaters.

Ergebnisse – Mischwasserentlastung Dresden



Mischwasserentlastung Dresden *Escherichia coli*



Trends:

- Resistenzniveau in der Entlastung ist entweder gleich oder geringer als im Ablauf der Kläranlage
- Resistenzniveau im Ablauf der Kläranlage entweder gleich oder größer als im Zulauf zur Kläranlage (einzige Ausnahme: FOX)

Wiederbelebung eines „alten“ Themas

APPLIED AND ENVIRONMENTAL MICROBIOLOGY, Dec 1982, p. 1395–1403
0099-2240/82/121395-09\$02.00/0
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Vol. 44, No. 6

R-Plasmid Transfer in a Wastewater Treatment Plant

PATRICK A. MACH[†] AND D. JAY GRIMES^{‡*}

River Studies Center and Department of Biology, University of Wisconsin-La Crosse, La Crosse, Wisconsin



FEMS Microbiology Ecology 48 (2003) 325–335



Detection of antibiotic-resistant bacteria and their resistance genes in wastewater, surface water, and drinking water biofilms

Thomas Schwartz ^{a,*}, Wolfgang Kohnen ^b, Bernd Jansen ^b, Ursula Obst ^a

Antibiotika , Resistenzen und Bakterien in Kläranlagen LUA NRW, 2006, Dr. Eichler

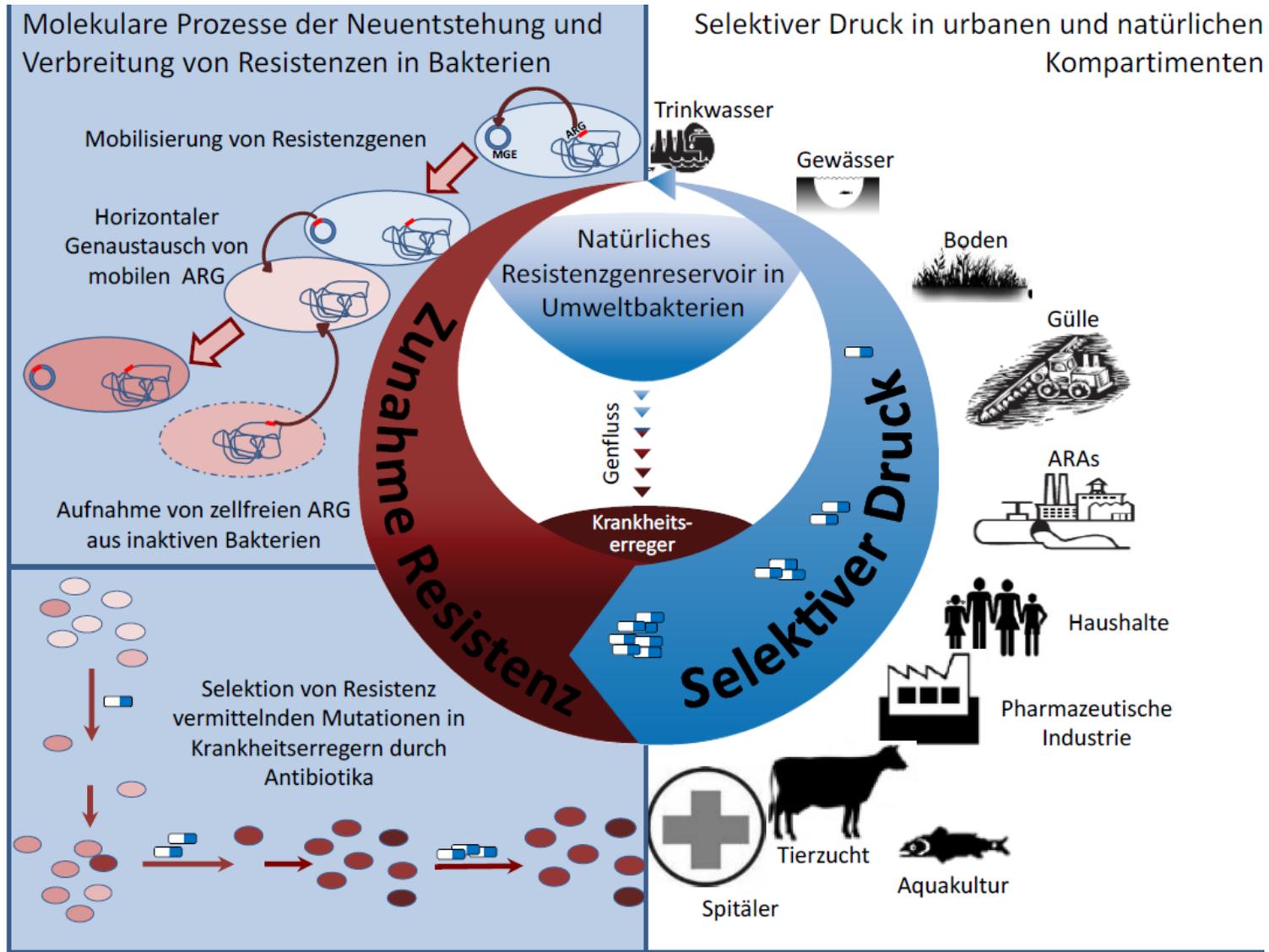
TZW-Schriftenreihe

Bedeutung von Antibiotikaresistenzen für die Rohwasserqualität: Vorkommen, Transport und natürliche Eliminationsprozesse
(2009) ISSN 1434-5765

Estimated total incidence of bloodstream infections with resistance phenotype (n per 100 000 population) and trend, 2019-2023, as well as the percentage change 2019-2023, by bacterial species and antimicrobial group/agent, Germany

Bacterial species	Antimicrobial group/agent	Estimated incidence* of isolates from bloodstream infections with resistance phenotype (n per 100 000 population)						
		2019	2020	2021	2022	2023	Trend 2019-2023 ^b	Change 2019-2023 (%) ^c
<i>E. coli</i>	Aminopenicillin (amoxicillin/ampicillin) resistance	50.64#	48.94#	44.66#	45.75#	47.05#	↓	-7.1
	Third-generation cephalosporin (cefotaxime/ceftriaxone/ceftazidime) resistance	12.02#	10.74#	9.07#	9.58#	10.23#	-	-14.9
	Carbapenem (imipenem/meropenem) resistance	0.01#	0.02#	0.04#	0.04#	0.05#	↑	+400.0
	Fluoroquinolone (ciprofloxacin/levofloxacin/ofloxacin) resistance	18.28#	17.14#	14.61#	15.40#	17.47#	-	-4.4
	Aminoglycoside (gentamicin/netilmicin/tobramycin) resistance ^d	8.53#	7.41#	5.28#	4.95#	5.33#	↓	-37.5
	Combined resistance to third-generation cephalosporins, fluoroquinolones, and aminoglycosides ^d	3.20#	2.71#	2.11#	1.80#	2.03#	↓	-36.6
<i>K. pneumoniae</i>	Third-generation cephalosporin (cefotaxime/ceftriaxone/ceftazidime) resistance	2.57#	2.38#	2.33#	2.32#	2.32#	-	-9.7
	Carbapenem (imipenem/meropenem) resistance	0.20#	0.11#	0.18#	0.23#	0.25#	-	+25.0
	Fluoroquinolone (ciprofloxacin/levofloxacin/ofloxacin) resistance	2.76#	2.54#	2.41#	2.61#	2.69#	-	-2.5
	Aminoglycoside (gentamicin/netilmicin/tobramycin) resistance ^d	1.52#	1.17#	0.93#	0.90#	0.86#	↓	-43.4
<i>P. aeruginosa</i>	Combined resistance to third-generation cephalosporins, fluoroquinolones, and aminoglycosides ^d	0.99#	0.77#	0.57#	0.65#	0.63#	↓	-36.4
	Piperacillin-tazobactam resistance	1.08#	1.12#	1.30#	1.31#	1.33#	↑	+23.1
	Ceftazidime resistance	0.94#	0.96#	1.04#	0.96#	0.98#	-	+4.3
	Carbapenem (imipenem/meropenem) resistance	1.21#	1.35#	1.46#	1.25#	1.27#	-	+5.0
	Fluoroquinolone (ciprofloxacin/levofloxacin) resistance	1.26#	1.03#	0.98#	0.93#	0.92#	↓	-27.0
	Aminoglycoside (gentamicin/netilmicin/tobramycin) resistance ^e	0.38#	0.17^#	0.17#	0.20#	0.16#	NA	-57.9
	Combined resistance to ≥3 antimicrobial groups (among piperacillin-tazobactam, ceftazidime, carbapenems, fluoroquinolones and aminoglycosides) ^e	0.58#	0.59^#	0.60^#	0.60#	0.60#	NA	+3.4
<i>Acinetobacter species</i>	Carbapenem (imipenem/meropenem) resistance	0.04#	0.07#	0.09#	0.08#	0.08#	-	+100.0
	Fluoroquinolone (ciprofloxacin/levofloxacin) resistance	0.10#	0.11#	0.12#	0.11#	0.11#	-	+10.0
	Aminoglycoside (gentamicin/netilmicin/tobramycin) resistance ^d	0.08#	0.09#	0.08#	0.09^#	0.06#	-	-25.0
	Combined resistance to carbapenems, fluoroquinolones and aminoglycosides ^d	0.03#	0.04^#	0.05#	0.05^#	0.04#	-	+33.3
<i>S. aureus</i>	MRSA ^f	3.56#	2.91#	2.64#	2.24#	2.44#	↓	-31.5
<i>S. pneumoniae</i>	Penicillin non-wild-type ^g	0.50#	0.29#	0.32#	0.44#	0.63#	-	+26.0
	Macrolide (azithromycin/darithromycin/erythromycin) resistance	0.67#	0.35#	0.27#	0.47#	0.65#	-	-3.0
	Combined penicillin non-wild-type and resistance to macrolides ^g	0.25#	0.10#	0.09#	0.20#	0.27#	-	+8.0
<i>E. faecalis</i>	High-level gentamicin resistance	1.25^#	1.38^#	1.33^#	1.37^#	1.40^#	-	+12.0
<i>E. faecium</i>	Vancomycin resistance	3.28#	3.18#	3.51#	2.71#	1.86#	↓	-43.3

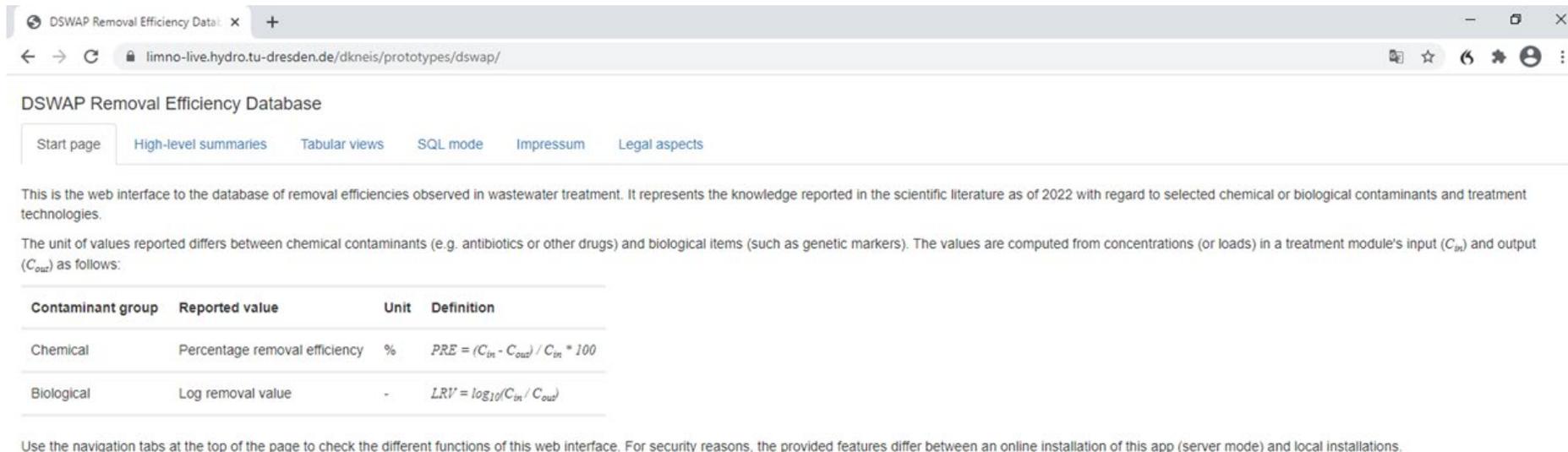
Resistente Bakterien, Gentransfer und Selektionsdruck



The End

Mögliche Methoden des AMR monitoring

Kontrollen - Behandlungstechnologien



DSWAP Removal Efficiency Database

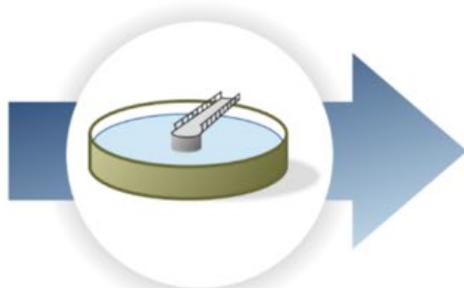
Start page | High-level summaries | Tabular views | SQL mode | Impressum | Legal aspects

This is the web interface to the database of removal efficiencies observed in wastewater treatment. It represents the knowledge reported in the scientific literature as of 2022 with regard to selected chemical or biological contaminants and treatment technologies.

The unit of values reported differs between chemical contaminants (e.g. antibiotics or other drugs) and biological items (such as genetic markers). The values are computed from concentrations (or loads) in a treatment module's input (C_{in}) and output (C_{out}) as follows:

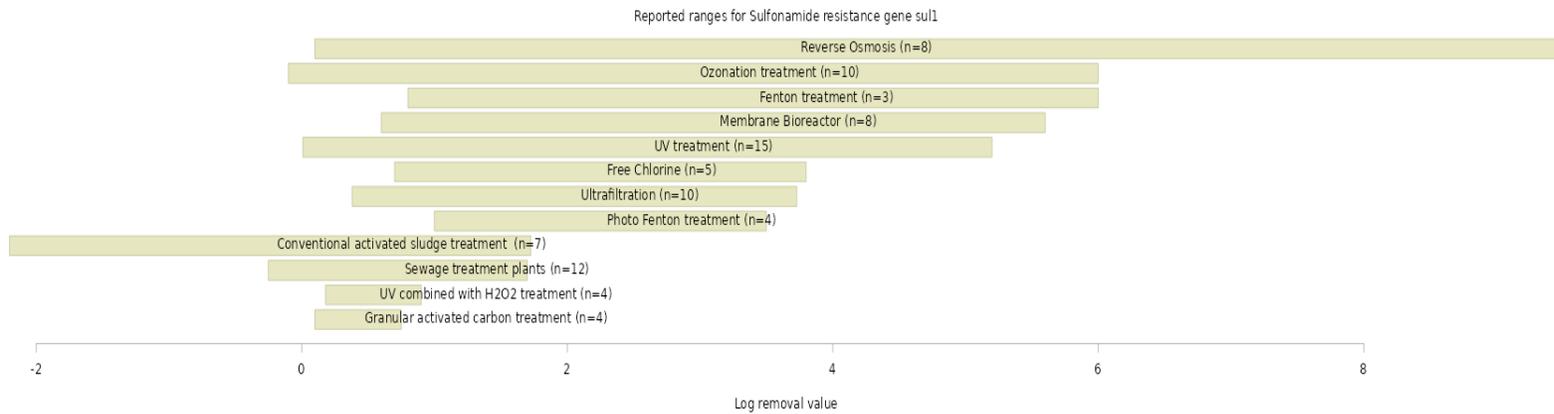
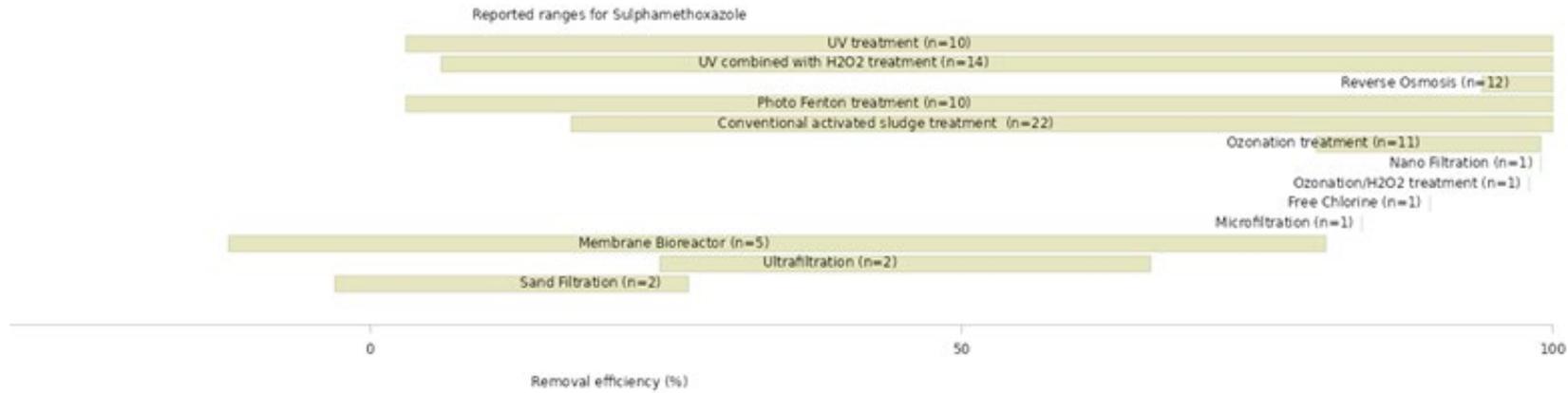
Contaminant group	Reported value	Unit	Definition
Chemical	Percentage removal efficiency	%	$PRE = (C_{in} - C_{out}) / C_{in} * 100$
Biological	Log removal value	-	$LRV = \log_{10}(C_{in} / C_{out})$

Use the navigation tabs at the top of the page to check the different functions of this web interface. For security reasons, the provided features differ between an online installation of this app (server mode) and local installations.



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The End