

Chronic Urticaria's Aetiology: Enlightening The Dark Side Of The Moon

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Abstract

Urticaria is characterized by wheals, erythema, and sometimes localized angioedema. It is defined as chronic (CU) when symptoms persist for more than 6 weeks; however the aetiology is incompletely understood. The aim of this study was to find causes and best treatments of CU.

For this purpose, personal and clinical data from 100 patients with CU were collected. The diagnostic work-up included stool parasite testing and the blood tests: IgG antiviral titres for EBV, CMV, HSV1, VZV and COVID-19; IgE for Anisakis; diamine oxidase (DAO); genetic testing for lactase deficiency; C3, C4; complete blood count; serum electrophoresis; anti-TG, anti-TPO, antinuclear and anti-dsDNA antibodies; P-ANCA, C-ANCA. Blood IgG antiviral Titers/Threshold Ratio (aVGt/TR) was evaluated in patients and 100 healthy controls (50 Male, 50 Female, 15–80 y.o.), divided into two groups, above and below 10 times the threshold value; statistical significance was assessed using the χ^2 test. Clinical outcomes, evaluated as changes in the UAS7 score before and after therapy, were categorized as: A) Healing; B) Improvement (decrease >70%); C) Unchanged (decrease <70%).

CU showed a M/F ratio = 0.52. Viral reinfections, identified by an aVGt/TR >10 ($p < 0.0005$), were the most frequent aetiological cause (71%), followed by autoimmunity (16%), anisakiasis (13%), histamine intolerance/DAO deficiency (3%), physical factors (2%), and idiopathic causes (2%). With targeted therapies, the patient-reported outcomes were: A) Healing 74%; B) Improvement 20%; C) Unchanged 6%.

CU aetiological diagnosis was achieved in 95% of cases. The term "spontaneous" chronic urticaria should be reserved for the refractory patients.

Key words: chronic urticaria aetiology, viruses, autoimmunity, skin mast cells

Introduction

Chronic urticaria has so far been classified (EAACI, WAO) on the basis of clinical criteria

-spontaneous and inducible- since attempts to identify its aetiology have often been unsuccessful (9). It is a disabling pathology, significantly impairing quality of life and interpersonal relationships (4,9). Recognized causes include cutaneous mastocytosis, mast cell activation syndrome, and autoimmune vasculitis; however, many syndromes presenting with chronic urticaria remain of unknown aetiology.

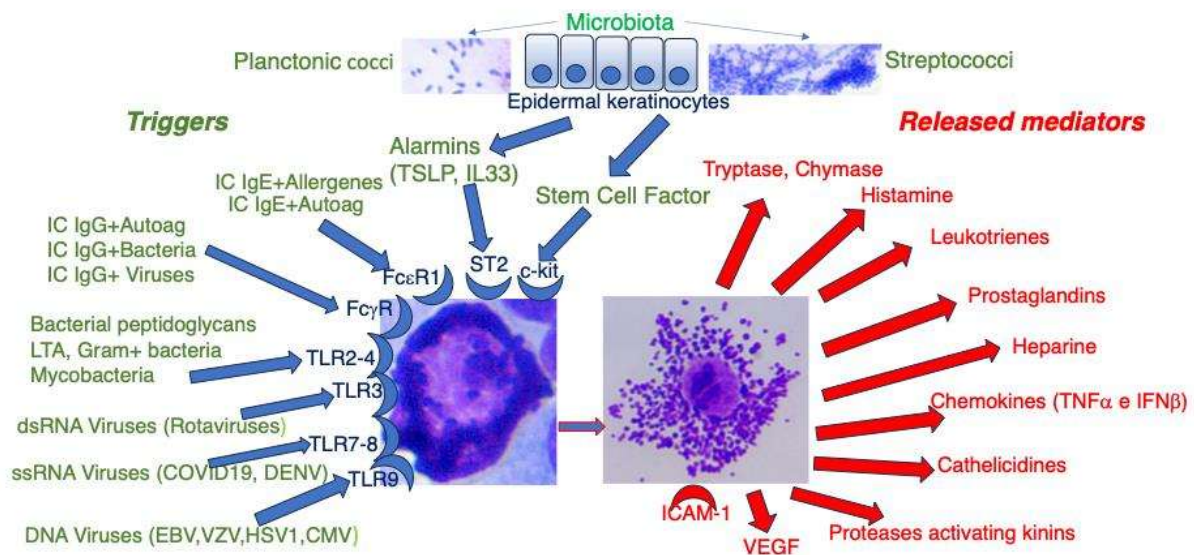
Symptoms such as pruritus, wheals, erythema, and angioedema derive from activation of cutaneous epithelial receptors (H1R and LTR), vasodilation, and increased vascular permeability.

They are attributable to the local release of mediators including histamine, leukotrienes, prostaglandins, heparin, chemokines (e.g., TNF α and IFN β), cathelicidins, and proteases, with activation of kinins (52,54). These mediators are stored in skin mast cells, which in connective tissues are typically equipped with both chymase and tryptase (MCTC), whereas mucosal mast cells contain mainly tryptase (MCT) (54).

Mast cells are effectors of both innate and adaptive immunity and can transition from a resting to an activated state in response to multiple triggers able to prompt degranulation with release of preformed mediators and synthesis of newly produced ones (2,5,6,8,32,55). Triggers include IgE–allergen immune complexes, IgE–autoantigen immune complexes, IgG–bacterial and IgG–viral immune complexes, RNA and DNA viruses, and bacterial peptidoglycans. Mast cells express pattern-recognition receptors, including Toll-like receptors (TLRs) and Pathogen Associated Molecular Patterns (PAMPs), whose engagement can induce activation and degranulation (27).

Skin keratinocytes challenged by viruses and bacteria (27) produce mediators including alarmins such as Thymic Stromal Lymphopoietin (TSLP) and IL-33, and growth factors such as Stem Cell Factor (SCF), which engages the c-kit receptor and results in mast cell proliferation and activation. IL-33 can induce synthesis and release of cytokines such as TNF α and IFN β , which exert antibacterial and antiviral effects, and it also increases expression of ICAM-1 (Fig. 1). On mucosal mast cells, ICAM-1 can facilitate engagement by rhinoviruses, enhancing cellular invasion (30). Circulating histamine is also produced by intestinal enterochromaffin cells during digestion (46,47), and in some cases its increase may be due to impaired catabolism by diamine oxidase (DAO), an enzyme produced by the epithelium covering the intestinal villi (7,42,43). This reduction may be primary or secondary and can lead to histamine intolerance. The aim of this study is therefore to identify the causes of chronic urticaria and the most effective treatments for a condition that remains only partially explained.

Fig1 Mastocytes degranulation's triggers in Chronic Urticaria.



Materials and Methods

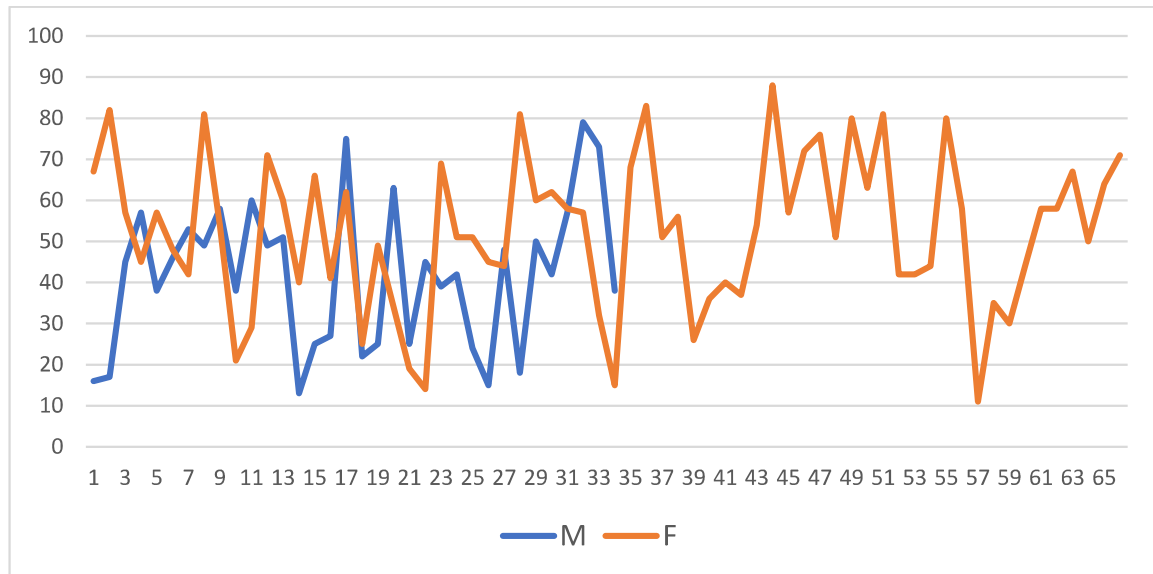
Subject to informed consent, computerized data were collected from 100 patients with chronic urticaria examined over the last 10 years, recording age, sex, identified aetiology, comorbidities such as gut microbiota subversion due to disaccharide intolerance, administered therapies, and patient-reported clinical outcome. The diagnostic pathway involved evaluation of IgG antiviral titres (EBV, CMV, HSV1, VZV, and COVID-19), specific IgE for Anisakis, DAO, genetic testing for lactase deficiency, C3, C4, complete blood count, serum electrophoresis, anti-TG and anti-TPO antibodies, antinuclear antibodies, anti-dsDNA antibodies, P-ANCA, C-ANCA, and stool parasite testing. Anti-viral IgG Titers/Threshold Ratio (aVGt/TR) values exceeding

10 times the threshold were distinguished from those below it, and the data were evaluated in patients and in 100 healthy controls (50 men, 50 women, 15–80 years old); the results were statistically analyzed using the χ^2 test. Clinical outcomes were assessed through the patient-reported UAS7 score before and after the administered therapies and were divided into three categories: A) Healing; B) Improvement (decrease >70%); C) Unchanged (decrease <70%).

Results

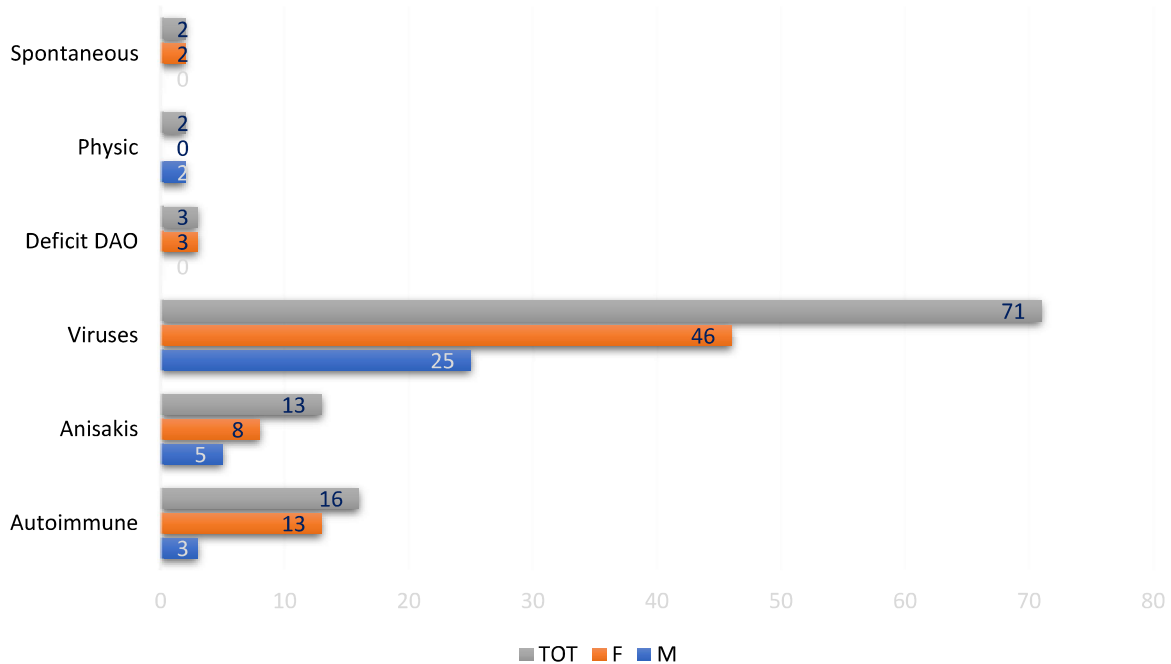
Chronic urticaria affected women approximately twice as frequently as men (M/F ratio = 0.52) and showed no age preference (11–81 years) (Fig. 2).

Fig.2 Age and Sex prevalence in 100 patients suffering from Chronic Urticaria



The aetiology included most frequently viral reinfections as responsible or jointly responsible (71%), followed by autoimmune causes (16%), anisakiasis (13%), DAO deficiency/histamine intolerance (3%), physical causes (2%), and spontaneous urticaria (2%) (Fig. 3).

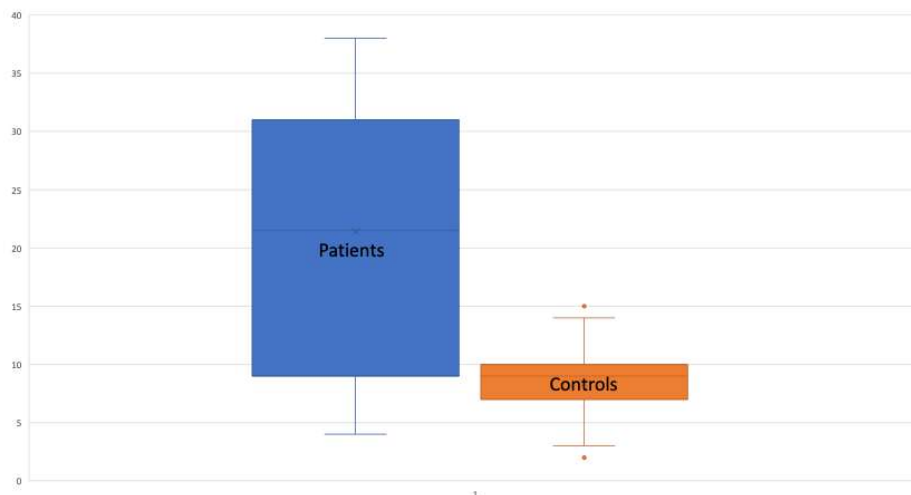
Fig 3 Chronic urticaria’s aetiology



Regarding the viral diagnostic work-up, the aVGt/TR in patients was significantly higher (more than 10-fold) compared to healthy controls (Fig. 4); the statistical analysis of the contingency table showed very high significance ($\chi^2 = 59.17$, $p < 0.0005$, Odds Ratio = 11.95, 99% confidence interval) (Tab.1).

Tab.1 Contingency table (aVGt/T Rt=antivirus IgG titers/Threshold Ratio)

aVGt/T Rt	Patients	Controls	TOT
>10	71	17	88
<10	29	83	112
TOT	100	100	200

Fig.4: Blood antiviral IgG titers/Threshold Ratio in CU Patients and Controls + SD.

Therefore, an IgG antiviral titre was considered a marker of recent viral reinfection/reactivation when the value exceeded approximately ten times the threshold value (8,19). Conversely, an aVGt/TR lower than 10-fold was attributed to immunological memory from past infection.

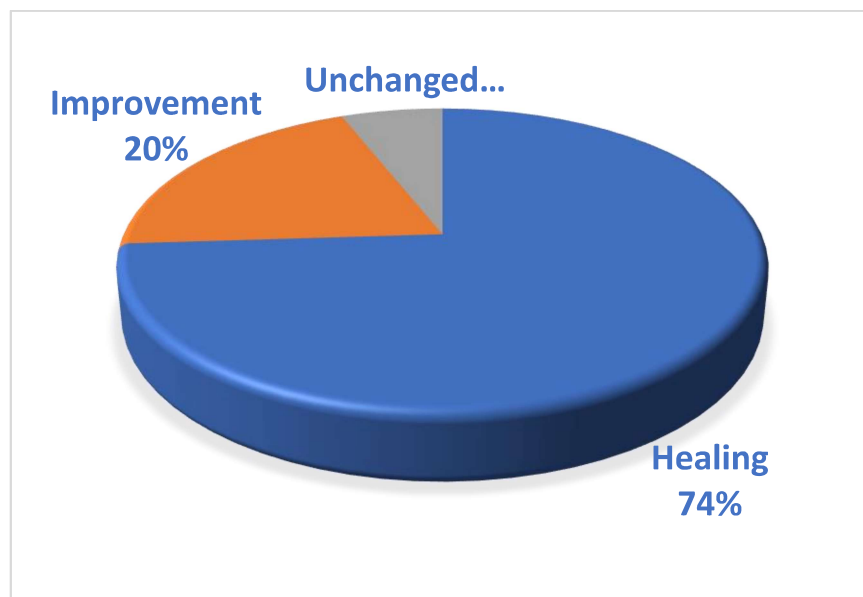
Therapies were aimed at treating symptoms (second-generation antihistamines, anti-leukotrienes, antioxidants, and low-dose corticosteroids only when necessary and for periods not exceeding 2–4 weeks), treating the identified aetiology (antivirals,

Fig.5 Global clinical outcomes in 100 patients suffering from Chronic Urticaria

anthelmintics, synthetic DAO, and small ozonized autohemotherapy), and managing relevant comorbidities (restoration of intestinal microbiota and treatment of underlying autoimmune diseases).

The patient-reported outcomes (Fig. 5) were:

- A) Healing 74%;
- B) Improvement 20%;
- C) Unchanged 6%.



Discussion

Chronic urticaria can be triggered by multiple factors that converge on a single effector responsible for symptoms: the cutaneous mast cell. Mast cells stem from bone-marrow-derived precursors that spread throughout the body; while their cytologic features are originally similar, their phenotype and mediator content diverge

according to tissue localization (27). In humans, mast cells resident in mucous membranes lining hollow organs exposed to the external environment (airways, gut, and genitourinary tract) contain mainly tryptase (MCT), whereas those resident in the skin and connective tissues of internal organs also contain chymase (MCTC).

These cells can be detected by optical microscopy using standardized cytological methods after classic May–Grünwald–

Giemsa staining (60). Vasodilation and neovessel proliferation with increased inflammation are induced by serine proteases, vascular endothelial growth factor (VEGF), other mitogens, and histamine released by mast cells (52,54).

Many triggers can unleash mast cell degranulation (Fig. 1): IgE–allergen immune complexes leading to classic type I reactions; IgE–autoantigen immune complexes (22,55); IgG–pathogen immune complexes; IgG–autoantigen immune complexes; viruses; and bacterial products such as peptidoglycans and lipoteichoic acid. These triggers engage different membrane structures, including receptors for alarmins (IL-33, Thymic Stromal Lymphopoietin -TSLP-) (63), Fc receptors, and pattern-recognition receptors (PAMPs and TLRs): FcγR for IgG immune complexes, FcεRI for IgE immune complexes, TSLPR, ST2 for IL-33, TLR2 and TLR4 for bacterial peptidoglycans and lipoteichoic acid, TLR3 for dsRNA viruses (e.g., rotaviruses), TLR7/8 for ssRNA viruses (e.g., COVID-19 and DENV), and TLR9 for DNA viruses (e.g., EBV, HSV, CMV, and VZV) (27).

Amplification of triggering effects may arise from keratinocytes stimulated by invading viruses and/or bacteria. Skin epithelial cells then release emergency signals such as alarmins (TSLP, IL-33) and growth factors such as SCF. SCF engages the mast cell c-kit receptor, inducing proliferation and degranulation, while IL-33 increases expression of membrane ICAM-1. ICAM-1 promotes intercellular adhesion and interactions; at mucosal surfaces it can also serve as a gateway for human rhinovirus (HRV), potentially increasing invasion (30). Increased production of VEGF enhances angiogenesis and local inflammation and has been associated with adverse prognostic features in oncology (53).

Viruses can also directly infect target cells, including mast cells. Regarding viral diagnostic work-up, it should be considered that viruses live and bud within parasitized cells, and most of their life cycle occurs in the cytoplasm. Therefore, viraemia may be transient, and serum polymerase chain reaction testing may be uninformative if sampling does not coincide with viral shedding into the bloodstream. Moreover, viruses may persist within circulating cells (e.g., lymphocytes), which can act as carriers to reach target tissues, acting as a sort of Trojan horse. Some reticuloendothelial-system-rich organs can act as reservoirs for virions; cytoplasmic proteolytic enzymes enable the partial digestion of viral capsid proteins, leading to viral transcription (61).

Another route used by viruses to invade cells is via neural axons, as in the case of shingles; the entry point in this case is the acetylcholine receptor (AChR), the same target of neurotoxins contained in some snake venoms. Viruses can cross this gateway bidirectionally, invading both neuronal axons and the cells at the neuromuscular junction, including mast cells.

In the skin, mast cells are located in the papillary dermis, near nerve fibres and around dermal blood vessels (62); therefore, they could plausibly contribute to local viral persistence and immune activation in chronic urticaria. Circulating antibodies against viruses are commonly detectable in blood, and their titre reflects infection history. After first exposure, IgM antibodies are produced and then replaced after about two weeks by IgG. Reinfections or reactivations may manifest mainly as an IgG increase. In immunocompetent subjects, memory T and B cells persist for about ten years; if reinfection occurs within this period, IgG synthesis can be faster and higher (8,13,18,26), and IgM synthesis does not

occur. Therefore, a virus-specific IgG titre less than about 10 times the threshold may reflect past immunological memory, whereas higher titres were interpreted as a marker of reinfection/reactivation, with greater values suggesting more recent infection. In our cohort, the ratio of virus-specific IgG titres to threshold was significantly higher (more than 10-fold) in patients with chronic viral urticaria (71%) than in healthy controls (17%). Monitoring the antiviral IgG titre/threshold ratio over time can help clinicians assess infection behaviour: a decreasing ratio indicates past infection, whereas a rising ratio suggests ongoing activity.

Regarding therapy, treating viruses may require antivirals, antioxidants, immunomodulators (e.g., lysozyme), and correction of factors associated with susceptibility to infections (e.g., restoration of intestinal microbiota in dysbiosis, correction of nutritional deficiencies, management of comorbidities). A case report described the efficacy of small ozonized autohemotherapy (intramuscular injection of the patient's blood exposed to an oxygen–ozone mixture), which resolved two cases of chronic urticaria associated with EBV and HSV1 (16). In this approach, exposure of blood to oxygen–ozone may inactivate pathogens because ozone has documented bactericidal and virucidal activity (57,58,59); the intramuscular injection of ozonated autologous blood, when performed according to validated protocols, may act as a form of autovaccine (64).

For gut parasites, including Anisakis, anthelmintic drugs such as mebendazole or albendazole are appropriate; numerous complications have been described in untreated anisakiasis (14,31,33–35), including intestinal bleeding (39), gastric cancer (36,37), intestinal ischemia (40), extraintestinal larval migration with respiratory manifestations (41), eosinophilic esophagitis (32), and hypereosinophilic syndrome (38).

After excluding autoimmune diseases, gut parasites, or viral causes, DAO deficiency may be detected. DAO is produced by intestinal villus epithelial cells and catabolizes histamine released by intestinal mast cells and enterochromaffin cells during digestion. In primary or secondary deficiency, circulating histamine may persist longer, potentially contributing to chronic urticaria; this clinical picture is often referred to as histamine intolerance (7,42,43,45). In such cases, it is appropriate to administer synthetic DAO and prescribe a low histamine-releasing diet. A randomized double-blind clinical trial showed synthetic DAO to be helpful in patients with chronic urticaria (45).

Moreover, failure to maintain the cold chain during storage of frozen foods (e.g., fish) can lead to a histaminergic syndrome mimicking anaphylaxis (Scombroid Fish Syndrome) (56), due to high histamine content resulting from bacterial histidine decarboxylase activity in improperly stored food.

Conclusions

In conclusion, mast cells in the skin and at mucosal surfaces act as advanced immune sentinels in barrier tissues. They are equipped with a wide range of membrane receptors for environmental cues and contain preformed mediators ready for release, while also being able to synthesize new ones when needed. The breadth of receptors expressed by mast cells can make them targets of pathogen-associated stimuli; in our cohort, viral infections were frequently associated with chronic urticaria. While host-protective, mast-cell-mediated responses may sometimes damage host tissues and contribute to trigger conditions such as chronic urticaria.

A structured diagnostic work-up combined with careful assessment of triggers able to activate skin mast cells allowed identification of aetiology in 95% of cases; targeted therapies achieved healing in three-quarters of patients and improvement in one-fifth. Only 6% of patients did not respond effectively; thus, the term “spontaneous” should arguably be reserved for these cases of chronic urticaria. Hospitalization and the use of biologic drugs limited to such refractory patients could involve significant cost savings for the National Health Service.

Compliance with Ethical Standards statements

The Author declares that there is no funding and no conflict of interest associated with this manuscript.

Impact Statement

Chronic spontaneous urticaria is often treated with expensive biologic drugs. By performing an appropriate diagnostic work-up and applying targeted treatments, it is possible to achieve substantial public cost savings for the National Health Service.

References

1. Ensina LF, Cusato-Ensina AP, Cardona R. Advances in the pathogenesis representing definite outcomes in chronic urticaria. *Curr Opin Allergy Clin Immunol*. 2019 Jun;19(3):193-197. doi: 10.1097/ACI.0000000000000519. PMID: 30676342.
2. Imbalzano E, Casciaro M, Quartuccio S, Minciullo PL, Cascio A, Calapai G, et al. Association between urticaria and virus infections: A systematic review. *Allergy Asthma Proc*. 2016 Jan-Feb;37(1):18-22. doi: 10.2500/aap.2016.37.3915. Epub 2015 Dec 3. PMID: 26637522.
3. Dreyfus DH. Autoimmune disease: A role for new anti-viral therapies? *Autoimmun Rev*. 2011 Dec;11(2):88-97. doi: 10.1016/j.autrev.2011.08.005. Epub 2011 Aug 18. PMID: 21871974.
4. Maurer M, Giménez-Arnau A, Ensina LF, Chu CY, Jaumont X, Tassinari P. Chronic urticaria treatment patterns and changes in quality of life: AWARE study 2-year results. *World Allergy Organ J*. 2020 Sep 12;13(9):100460. doi: 10.1016/j.waojou.2020.100460. PMID: 32983330; PMCID: PMC7493083.
5. Sachdeva M, Gianotti R, Shah M, Bradanini L, Tosi D, Veraldi S, et al. Cutaneous manifestations of COVID-19: Report of three cases and a review of literature. *J Dermatol Sci*. 2020 May;98(2):75-81. doi: 10.1016/j.jdermsci.2020.04.011. Epub 2020 Apr 29. PMID: 32381430; PMCID: PMC7189855.
6. Kocatürk, E.; Muñoz, M.; Elieh-Ali-Komi, D.; Criado, P.R.; Peter, J.; Kolkhir, et al. How Infection and Vaccination Are Linked to Acute and Chronic Urticaria: A Special Focus on COVID-19. *Viruses* 2023, 15, 1585. <https://doi.org/10.3390/v15071585>
7. Comas-Basté, O.; Sánchez-Pérez, S.; Veciana-Nogués, M.T.; Latorre-Moratalla, M.; Vidal-Carou, M.d.C. Histamine Intolerance: The Current State of the Art. *Biomolecules* 2020, 10, 1181. <https://doi.org/10.3390/biom10081181>
8. Dreyfus DH. Serological evidence that activation of ubiquitous human herpesvirus-6 (HHV-6) plays a role in chronic idiopathic/spontaneous urticaria (CIU). *Clin Exp Immunol*. 2016 Feb;183(2):230-8. doi: 10.1111/cei.12704. Epub 2015 Nov 24. PMID: 26361716; PMCID: PMC4711162.
9. Zuberbier T, Abdul Latiff AH, Abuzakouk M, Aquilina S, Asero R, Baker D, et al. The international EAACI/GA²LEN/EuroGuiDerm/APAAACI guideline for the definition, classification, diagnosis, and management of urticaria. *Allergy*. 2022 Mar;77(3):734-766. doi: 10.1111/all.15090. Epub 2021 Oct 20. PMID: 34536239.
10. Lemus, Y.B., Martínez, G.A., Lugo, L.P. et al. Author Correction: Gene profiling of Epstein-Barr Virus and human endogenous retrovirus in peripheral blood mononuclear cells of SLE patients: immune response implications. *Sci Rep* 14, 22032 (2024). <https://doi.org/10.1038/s41598-024-73833->
11. Hoeggerl AD, Nunhofer V, Lauth W, Badstuber N, Held N, Zimmermann G, et al. Epstein-Barr virus reactivation is not causative for post-COVID-19-syndrome in individuals with asymptomatic or mild SARS-CoV-2 disease course. *BMC Infect Dis*. 2023 Nov 15;23(1):800. doi: 10.1186/s12879-023-08820-w. PMID: 37968601; PMCID: PMC10652630.
12. Lieberman PM. Virology. Epstein-Barr virus turns 50. *Science*. 2014 Mar 21;343(6177):1323-5. doi: 10.1126/science.1252786. PMID: 24653027; PMCID: PMC4581426.
13. Andersen O, Ernberg I, Hedström AK. Treatment Options for Epstein-Barr Virus-Related Disorders of the Central Nervous System. *Infect Drug Resist*. 2023 Jul 13;16:4599-4620. doi: 10.2147/IDR.S375624. PMID: 37465179; PMCID: PMC10351589.
14. Cherrez Ojeda I, Vanegas E, Felix M, Mata V, Cherrez S, Simancas-Racines D, et al. Etiology of chronic urticaria: the Ecuadorian experience. *World Allergy Organ J*. 2018 Jan 3;11(1):1. doi: 10.1186/s40413-017-0181-0. PMID: 29308115; PMCID: PMC5753451.
15. Widney D, Boscardin WJ, Kasravi A, Martínez-Maza O. Expression and function of CD28 on Epstein-Barr virus-positive B cell lines and AIDS-associated non-Hodgkin's lymphoma cell lines. *Tumour Biol*. 2003 Mar-Apr;24(2):82-93. doi: 10.1159/000071081. PMID: 12853703.
16. Manfredi G., Apuzzo D.: Successful Ozone Treatment of EBV and HSV-Related Viral Urticaria *Frontiers in Medical Case Reports* March 2020 , 01 (02): 1-6
17. Albanese M, Tagawa T, Hammerschmidt W. Strategies of Epstein-Barr virus to evade innate antiviral immunity of its human host. *Front Microbiol*. 2022 Jul 22;13:955603. doi: 10.3389/fmicb.2022.955603. PMID: 35935191; PMCID: PMC9355577.
18. Mareri A, Adler SP, Nigro G. Herpesvirus-associated acute urticaria: an age matched case-control study. *PLoS One*. 2013 Dec 27;8(12):e85378. doi: 10.1371/journal.pone.0085378. PMID: 24386470; PMCID: PMC3874042.
19. Harada S, Kamata Y, Ishii Y, Eda H, Kitamura R, Obayashi M, et al. Maintenance of serum immunoglobulin G antibodies to Epstein-Barr virus (EBV) nuclear antigen 2 in healthy individuals from different age groups in a Japanese population with a high childhood incidence of asymptomatic primary EBV infection. *Clin Diagn Lab Immunol*. 2004 Jan;11(1):123-30. doi: 10.1128/cdli.11.1.123-130.2004. PMID: 14715558; PMCID: PMC321344.
20. Rudenko M. The role of infection and autoimmunity in Urticaria and angioedema as a common entity. *EMJ Allergy*

- Immunol 2021 (6): 79-85.
21. James JA, Robertson JM. Lupus and Epstein-Barr. *Curr Opin Rheumatol.* 2012 Jul;24(4):383-8. doi: 10.1097/BOR.0b013e3283535801. PMID: 22504579; PMCID: PMC3562348.
 22. Charles N, Kortekaas-Krohn I, Kocaturk E, Scheffel J, Altrichter S, Steinert C, et al. Autoreactive IgE: Pathogenic role and therapeutic target in autoimmune diseases. *Allergy.* 2023 Dec;78(12):3118-3135. doi: 10.1111/all.15843. Epub 2023 Aug 9. PMID: 37555488.
 23. Sun Q, Li W, She R, Wang D, Han D, Li R, et al. Evidence for a role of mast cells in the mucosal injury induced by Newcastle disease virus. *Poult Sci.* 2009 Mar;88(3):554-61. doi: 10.3382/ps.2008-00468. PMID: 19211524.
 24. Burke SM, Issekutz TB, Mohan K, Lee PW, Shmulevitz M, Marshall JS. Human mast cell activation with virus-associated stimuli leads to the selective chemotaxis of natural killer cells by a CXCL8-dependent mechanism. *Blood.* 2008 Jun 15;111(12):5467-76. doi: 10.1182/blood-2007-10-118547. Epub 2008 Apr 18. PMID: 18424663.
 25. Shirato K, Taguchi F. Mast cell degranulation is induced by A549 airway epithelial cell infected with respiratory syncytial virus. *Virology.* 2009 Mar 30;386(1):88-93. doi: 10.1016/j.virol.2009.01.011. Epub 2009 Feb 4. PMID: 19195674.
 26. De Paschale M, Clerici P. Serological diagnosis of Epstein-Barr virus infection: Problems and solutions. *World J Virol.* 2012 Feb 12;1(1):31-43. doi: 10.5501/wjv.v1.i1.31. PMID: 24175209; PMCID: PMC3782265.
 27. Igawa S, Di Nardo A. Skin microbiome and mast cells. *Transl Res.* 2017 Jun;184:68-76. doi: 10.1016/j.trsl.2017.03.003. Epub 2017 Mar 23. PMID: 28390799; PMCID: PMC5538027.
 28. Liu Y, de Waal Malefyt R, Briere F, Parham C, Bridon JM, Banchereau J, et al. The EBV IL-10 homologue is a selective agonist with impaired binding to the IL-10 receptor. *J Immunol.* 1997 Jan 15;158(2):604-13. PMID: 8992974.
 29. Syenina A, Saron WAA, Jagaraj CJ, Bibi S, Arock M, Gubler DJ, et al. Th1-Polarized, Dengue Virus-Activated Human Mast Cells Induce Endothelial Transcriptional Activation and Permeability. *Viruses.* 2020 Dec 2;12(12):1379. doi: 10.3390/v12121379. PMID: 33276578; PMCID: PMC7761533.
 30. Akoto C, Willis A, Banas CF, Bell JA, Bryant D, Blume C, et al. IL-33 Induces an Antiviral Signature in Mast Cells but Enhances Their Permissiveness for Human Rhinovirus Infection. *Viruses.* 2022 Nov 1;14(11):2430. doi: 10.3390/v14112430. PMID: 36366528; PMCID: PMC9699625.
 31. Dias Fde J, Clemente SC, Knoff M. Nematoides anisacuídeos e cestoides Trypanorhyncha de importância em saúde pública em Aluterus monoceros (Linnaeus, 1758) no Estado do Rio de Janeiro, Brasil [Larvae of Anisakidae nematodes and Trypanorhyncha cestodes of public health importance in Aluterus monoceros (Linnaeus, 1758) in Rio de Janeiro State, Brazil]. *Rev Bras Parasitol Vet.* 2010 Apr-Jun;19(2):94-7. Portuguese. PMID: 20624345.
 32. Bircher AJ, Gysi B, Zenklusen HR, Aerni R. Eosinophile Oesophagitis assoziiert mit rezidivierender Urtikaria: Steckt da der Wurm Anisakis simplex drin? [Eosinophilic esophagitis associated with recurrent urticaria: is the worm Anisakis simplex involved?]. *Schweiz Med Wochenschr.* 2000 Nov 25;130(47):1814-9. German. PMID: 11130147.
 33. Carlin AF, Abeles S, Chin NA, Lin GY, Young M, Vinetz JM. Case Report: A Common Source Outbreak of Anisakidosis in the United States and Postexposure Prophylaxis of Family Collaterals. *Am J Trop Med Hyg.* 2018 Nov;99(5):1219-1221. doi: 10.4269/ajtmh.18-0586. PMID: 30226150; PMCID: PMC6221231.
 34. Nonković, D.; Tešić, V.; Šimat, V.; Karabuva, S.; Medić, A.; Hrabar, J. Anisakidae and Anisakidosis: A Public Health Perspective. *Pathogens* **2025**, *14*, 217. <https://doi.org/10.3390/pathogens14030217>
 35. Adroher, F.J.; Morales-Yuste, M.; Benítez, R. Anisakiasis and Anisakidae. *Pathogens* **2024**, *13*, 148. <https://doi.org/10.3390/pathogens13020148>
 36. Sakurai E, Okubo M, Tsutsumi Y, Shibata T, Tahara T, Kiriya Y, et al. A case of chronic gastric anisakiasis coexisting with early gastric cancer. *Fujita Med J.* 2023 May;9(2):163-169. doi: 10.20407/fmj.2022-010. Epub 2022 Oct 28. PMID: 37234391; PMCID: PMC10206891.
 37. Tsutsumi Y, Fujimoto Y. Early gastric cancer superimposed on infestation of an Anisakis-like larva: a case report. *Tokai J Exp Clin Med.* 1983 Jul;8(3):265-73. PMID: 6686894.
 38. Nicola S, Borrelli R, Ridolfi I, Lo Sardo L, Negrini S, Fornero M, et al. From hypereosinophilia to hypereosinophilic syndrome: real-world application of a two-tailed approach for HES diagnosis. *Front Immunol.* 2026 Jan 6;16:1735131. doi: 10.3389/fimmu.2025.1735131. PMID: 41567200; PMCID: PMC12815814.
 39. Filauro M, Rollandi GA, Cassola G, Quilici P, Angelini G, Belli F, et al. Gastrointestinal bleeding due to suspected anisakiasis: challenging differential diagnosis for a rare disease. *Updates Surg.* 2011 Sep;63(3):213-7. doi: 10.1007/s13304-011-0055-x. Epub 2011 Mar 29. PMID: 21445646.
 40. Shibata K, Yoshida Y, Miyaoka Y, Emoto S, Kawai T, Kobayashi S, et al. Intestinal anisakiasis with severe intestinal ischemia caused by extraluminal live larvae: a case report. *Surg Case Rep.* 2020 Oct 1;6(1):253. doi: 10.1186/s40792-020-01033-2. PMID: 33001287; PMCID: PMC7530153.
 41. Saito W, Kawakami K, Kuroki R, Matsuo H, Oishi K, Nagatake T. Pulmonary anisakiasis presenting as eosinophilic pleural effusion. *Respirology.* 2005 Mar;10(2):261-2. doi: 10.1111/j.1440-1843.2005.00643.x. PMID: 15823197.
 42. Weidenhiller M, Layritz C, Hagel AF, Kuefner M, Zopf Y, Raithel M. Histaminintoleranz-Syndrom (HIS): Vielfalt der Mechanismen von physiologischer, pathophysiologischer und toxischer Wirkung und deren Unterscheidung [Histamine intolerance syndrome (HIS): plethora of physiological, pathophysiological and toxic mechanisms and their differentiation]. *Z Gastroenterol.* 2012 Dec;50(12):1302-9. German. doi: 10.1055/s-0032-1325487. Epub 2012 Dec 7. PMID: 23225559.
 43. Hanusková E, Plevková J. Histaminová intolerancia [Histamine intolerance]. *Cesk Fysiol.* 2013;62(1):26-33. Slovak. PMID: 23821960.
 44. Kettner L, Seitzl I, Fischer L. Recent advances in the application of microbial diamine oxidases and other

- histamine-oxidizing enzymes. *World J Microbiol Biotechnol.* 2022 Oct 8;38(12):232. doi: 10.1007/s11274-022-03421-2. PMID: 36208352; PMCID: PMC9547800.
45. Yacoub MR, Ramirez GA, Berti A, Mercurio G, Breda D, Saporiti N, Bet al. Diamine Oxidase Supplementation in Chronic Spontaneous Urticaria: A Randomized, Double-Blind Placebo-Controlled Study. *Int Arch Allergy Immunol.* 2018;176(3-4):268-271. doi: 10.1159/000488142. Epub 2018 Apr 26. PMID: 29698966.
46. Mohan BP, Aravindan KP. Enterochromaffin Cells and Mast Cells in Acute Appendicitis. *J Lab Physicians.* 2020 Aug;12(2):141-146. doi: 10.1055/s-0040-1716476. Epub 2020 Sep 1. PMID: 32905235; PMCID: PMC7466542.
47. Yakabi K, Kawashima J, Kato S. Ghrelin and gastric acid secretion. *World J Gastroenterol.* 2008 Nov 7;14(41):6334-8. doi: 10.3748/wjg.14.6334. PMID: 19009648; PMCID: PMC2766114.
48. Bechi P, Romagnoli P, Panula P, Dei R, Bacci S, Amorosi A, et al. Gastric mucosal histamine storing cells. Evidence for different roles of mast cells and enterochromaffin-like cells in humans. *Dig Dis Sci.* 1995 Oct;40(10):2207-13. doi: 10.1007/BF02209008. PMID: 7587791.
49. Heidarzadeh-Asl S, Maurer M, Kiani A, Atiakshin D, Stahl Skov P, Elieh-Ali-Komi D. Novel insights on the biology and immunologic effects of histamine: A road map for allergists and mast cell biologists. *J Allergy Clin Immunol.* 2025 Apr;155(4):1095-1114. doi: 10.1016/j.jaci.2024.12.1081. Epub 2024 Dec 27. PMID: 39734034.
50. Melgarejo E, Medina MA, Sánchez-Jiménez F, Urdiales JL. Targeting of histamine producing cells by EGCG: a green dart against inflammation? *J Physiol Biochem.* 2010 Sep;66(3):265-70. doi: 10.1007/s13105-010-0033-7. Epub 2010 Jul 22. PMID: 20652470.
51. Masini E, Di Bello MG, Raspanti S, Ndisang JF, Baronti R, Cappugi P, et al. The role of histamine in platelet aggregation by physiological and immunological stimuli. *Inflamm Res.* 1998 May;47(5):211-20. doi: 10.1007/s000110050319. Erratum in: *Inflamm Res.* 2013 Feb;62(2):249. Fomusi Ndisang, J [corrected to Ndisang, J F]. PMID: 9657253.
52. Katunuma N, Kido H. Biological functions of serine proteases in mast cells in allergic inflammation. *J Cell Biochem.* 1988 Dec;38(4):291-301. doi: 10.1002/jcb.240380408. PMID: 2467915.
53. Ozdemir O. Immunosurveillance function of human mast cell? *World J Gastroenterol.* 2005 Nov 28;11(44):7054-6. doi: 10.3748/wjg.v11.i44.7054. PMID: 16437618; PMCID: PMC4717056.
54. Caughey GH. Mast cell tryptases and chymases in inflammation and host defense. *Immunol Rev.* 2007 Jun;217:141-54. doi: 10.1111/j.1600-065X.2007.00509.x. PMID: 17498057; PMCID: PMC2275918.
55. Kolkhir P, Muñoz M, Asero R, Ferrer M, Kocatürk E, Metz M, et al. Autoimmune chronic spontaneous urticaria. *J Allergy Clin Immunol.* 2022 Jun;149(6):1819-1831. doi: 10.1016/j.jaci.2022.04.010. PMID: 35667749.
56. Gingras V, Marois L. Case of scombroid syndrome mimicking an anaphylactic fish allergy. *Int J Emerg Med.* 2025 Jul 3;18(1):122. doi: 10.1186/s12245-025-00930-3. PMID: 40610892; PMCID: PMC12224635.
57. Larini A, Bocci V. Effects of ozone on isolated peripheral blood mononuclear cells. *Toxicol In Vitro.* 2005 Feb;19(1):55-61. doi: 10.1016/j.tiv.2004.06.007. PMID: 15582356.
58. Orta de Velásquez MT, Yáñez Noguez I, Casasola Rodríguez B, Román Román PI. Effects of ozone and chlorine disinfection on VBNC *Helicobacter pylori* by molecular techniques and FESEM images. *Environ Technol.* 2017 Mar;38(6):744-753. doi: 10.1080/09593330.2016.1210680. Epub 2016 Jul 26. PMID: 27432258.
59. Hogard S, Pearce R, Yetka K, Gonzalez R, Bott C. Virus inactivation in low ozone exposure water reuse applications. *Water Res.* 2024 Jun 1;256:121536. doi: 10.1016/j.watres.2024.121536. Epub 2024 Mar 28. PMID: 38631238.
60. Macchi A, Gelardi M, Landi M, Gramellini G, Giancaspro R, Nappi L, et al. Standardization of Nasal Cytology: An Expert-based Delphi Consensus. *Curr Allergy Asthma Rep.* 2026 Feb 21;26(1):15. doi: 10.1007/s11882-026-01256-7. PMID: 41721147; PMCID: PMC12923426.
61. Mandell, Douglas and Bennet's: Principle and practice of infectious diseases. Churchill Livingstone Inc., New York, 1995, 1314-1324
62. Criado PR, Jardim Criado RF, Ianhez M, Miot HA. Chronic pruritus: a narrative review. *An Bras Dermatol.* 2025 May-Jun;100(3):487-519. doi: 10.1016/j.abd.2024.09.008. Epub 2025 May 3. PMID: 40320333; PMCID: PMC12234205.
63. Varricchi G, Pecoraro A, Marone G, Criscuolo G, Spadaro G, Genovese A, et al. Thymic Stromal Lymphopoietin Isoforms, Inflammatory Disorders, and Cancer. *Front Immunol.* 2018 Jul 13;9:1595. doi: 10.3389/fimmu.2018.01595. PMID: 30057581; PMCID: PMC6053489.
64. Bocci, V., Zanardi, I., Michaeli, D., & Travagli, V. Mechanisms of action and chemical-biological interactions between ozone and body compartments: a critical appraisal of the different administration routes. *Current drug therapy* 2009, 4(3), 159-173.