

OPTIMIZING SEDATION IN FELINE BLOOD DONATION: THE ROLE OF ORAL GABAPENTIN PREMEDICATION

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INTRODUCTION

Sedation is often necessary to ensure the well-being of the animals and the safety of the blood donation. Oral gabapentin is increasingly used as premedication due to its anxiolytic and mild sedative effects, and also low incidence of adverse effects. However, evidence regarding its impact on IV sedation requirements in feline blood donor programs is limited.

This study aimed to evaluate the relation between gabapentin premedication and the required dose of IV sedation.

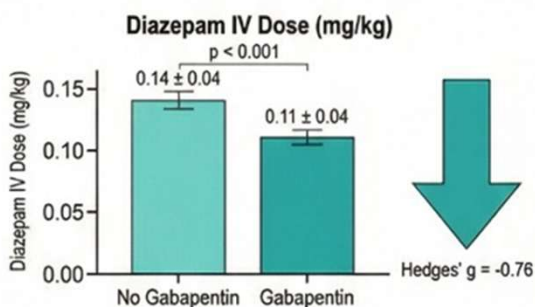
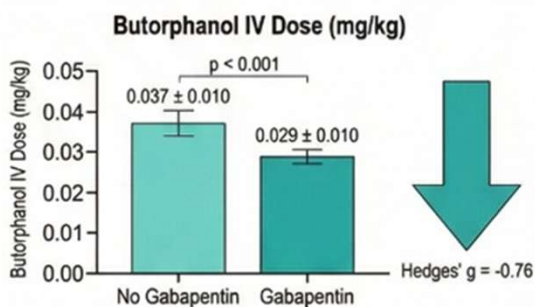
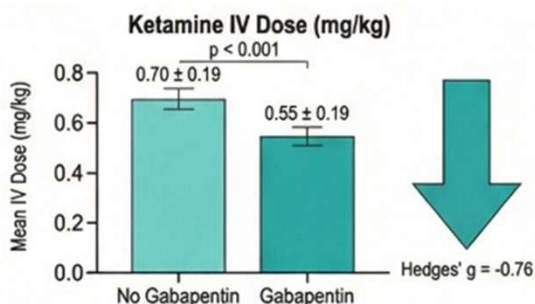
METHODOLOGY

Donation events were divided into two groups: cats receiving gabapentin premedication (100 mg/cat, administered 90 minutes before donation; n = 272) and cats not premedicated (n = 1,828). The recorded IV sedative dose in each donation event corresponded to the total administered dose, including any additional boluses when required. Intravenous doses (mg/kg) of ketamine, butorphanol, and diazepam (always used in combination) were compared between the two groups using independent-samples t-tests with Welch's correction. All tests were two-tailed.

DATA Feb 2025 - Jan 2026

Retrospective study 2,100 donations 1,357 sedated cats

RESULTS



STUDY POPULATION

52% Female
 64% Domestic Shorthair
 Mean age 4 years
 Mean weight 4,9 Kg



PREMEDICATION

With Gabapentin Premedication (n=272, 100mg PO, 90 min before)

No Gabapentin (n=1,828)

OUTCOME

IV Sedative dose (Ketamine, Butorphanol, Diazepam)

compared using t-tests (Welch's correction)

CONCLUSION

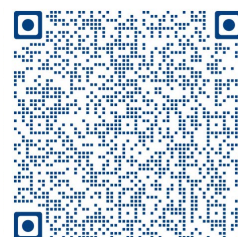
CONCLUSION

Oral gabapentin premedication was associated with a significant reduction in the required IV doses of all evaluated sedative agents.

CONTEXT & LIMITATIONS

Overall sedation need depends on donor reactivity. The potential bias of clinician-driven dose adjustments in the gabapentin group is considered unlikely.

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Introduction:

Sedation is often necessary to ensure the well-being of the animals and the safety of the blood donation procedure.

Oral gabapentin is increasingly used as premedication due to its anxiolytic and mild sedative effects, and also low incidence of adverse effects. However, evidence regarding its impact on IV sedation requirements in feline blood donor programs is limited. This study aimed to evaluate the relationship between gabapentin premedication and the required dose of IV sedation.

Methods:

A retrospective analysis was performed on 2,100 blood donations from 1,357 indoor, privately owned cats between February 2025 and January 2026. The study population consisted predominantly of female (52%) and domestic shorthair (64%) cats, with a mean age of 4 years (range between 1 and 8 years) and a mean body weight of 4.9 kg (range between 3 and 9 kg). Donation events were divided into two groups: cats receiving gabapentin premedication (100 mg/cat, administered 90 minutes before donation; n = 272) and cats not receiving gabapentin (n = 1,828). The recorded IV sedative dose in each donation event corresponded to the total administered dose, including any additional boluses when required. Intravenous doses (mg/kg) of ketamine, butorphanol, and diazepam were compared between the two groups using independent-samples t-tests with Welch's correction. All tests were two-tailed.

Results:

Oral gabapentin premedication was associated with a significant reduction in the required IV doses of all evaluated sedative agents. The mean (\pm SD) ketamine dose was significantly lower in the gabapentin group (0.55 ± 0.19 mg/kg) than in the non-gabapentin group (0.70 ± 0.19 mg/kg; $p < 0.001$). Similarly, the mean butorphanol dose decreased (from 0.029 ± 0.010 to 0.037 ± 0.010 mg/kg; $p < 0.001$), as did the mean diazepam dose (from 0.11 ± 0.04 to 0.14 ± 0.04 mg/kg; $p < 0.001$). Effect sizes were large for all comparisons (Hedges' $g = -0.76$; 95% CI, -0.89 to -0.63).

Conclusion:

Oral gabapentin premedication was associated with lower IV doses of sedatives during feline blood donation. Overall sedation need depends on donor reactivity during the donation, meaning the bias of potential clinician-driven IV sedatives dose adjustments in gabapentin group is unlikely.

PREVALENCE OF *DIROFILARIA IMMITIS* IN A POPULATION OF FELINE BLOOD DONORS

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INTRODUCTION

Dirofilaria immitis, the primary causative agent of feline heartworm disease, is a zoonotic, mosquito-borne filarioid with a worldwide distribution. While felines contribute to a lesser extent to the transmission of parasites compared to canines, infections do occur and are frequently underdiagnosed due to the low burden of parasites, the transient or absent nature of microfilaremia, and the nonspecific or subclinical nature of clinical signs. The diagnosis is further complicated by delayed or inconsistent antigenemia, particularly in cases of immature or unisexual infections. Given the potential for *D. immitis*-infected donors to pose diagnostic, vector, and health risks, screening and prophylaxis are recommended in endemic areas.

The objective of this study was to ascertain the prevalence of heartworm antigen in clinically healthy cats enrolled in a feline blood donor program.

CONCLUSION

During the study period, no evidence of circulating *Dirofilaria immitis* antigen was detected among clinically healthy cats enrolled in this feline blood donor program.

Notwithstanding the 90.2% test sensitivity, the present study indicates an exceedingly low prevalence of detectable heartworm infection within the specified population.

However, given the known limitations of antigen testing in cats, particularly in cases of low parasite burden or immature infections, the absence of positive results should be interpreted with caution.



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METHODOLOGY

A retrospective observational analysis was conducted using data from a feline blood donor program.



FELINE BLOOD DONOR PROGRAM 978 healthy cats	1,210 SAMPLES Feb-Jul 2025	SNAP Feline Triple Test (IDEXX)® Sensitivity 90.0%, Specificity 100%	DIROFILARIA IMMITIS Antigen detection
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RESULTS

Of the 1,210 samples that were examined, none exhibited a positive result for the presence of *Dirofilaria immitis* antigen.

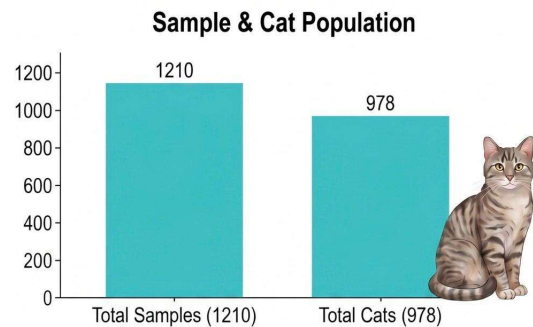


Figure 1. Sample & Cat Population

Dirofilaria immitis Antigen Test Results

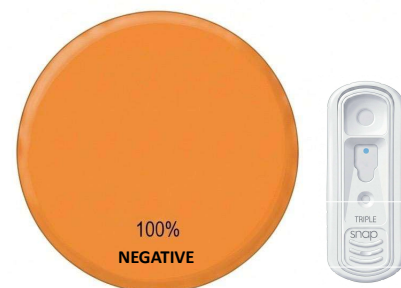


Figure 2. *Dirofilaria immitis* Test Results

PREVALENCE OF *DIROFILARIA IMMITIS* IN A POPULATION OF FELINE BLOOD DONORS

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Introduction:

Feline heartworm infection is characterized by the presence of migrating larvae or adult *Dirofilaria* spp. in host tissues or the pulmonary vasculature. *Dirofilaria immitis*, the primary causative agent of feline heartworm disease, is a zoonotic, mosquito-borne filarioid with a worldwide distribution. While felines contribute to a lesser extent to the transmission of parasites compared to canines, infections do occur and are frequently underdiagnosed due to the low burden of parasites, the transient or absent nature of microfilaremia, and the nonspecific or subclinical nature of clinical signs. The diagnosis is further complicated by delayed or inconsistent antigenemia, particularly in cases of immature or unisexual infections. Given the potential for *D. immitis*-infected donors to pose diagnostic, vector, and health risks, screening and prophylaxis are recommended in endemic areas. The objective of this study was to ascertain the prevalence of heartworm antigen in clinically healthy cats enrolled in a feline blood donor program.

Methods:

A retrospective observational analysis was conducted using data from a feline blood donor program. The study population comprised clinically healthy cats, who underwent screening using the SNAP[®] Feline Triple Test (sensitivity 90.2%, specificity 100%). The primary focus of this test was the detection of *Dirofilaria immitis* antigen. A total of 1,210 samples were collected from 978 cats between February and July of 2025 for the purpose of analysis.

Results:

Of the 1,210 samples that were examined, none exhibited a positive result for the presence of *Dirofilaria immitis* antigen.

Conclusions:

During the study period, no evidence of circulating *Dirofilaria immitis* antigen was detected among clinically healthy cats enrolled in this feline blood donor program. Notwithstanding the 90.2% test sensitivity, the present study indicates an exceedingly low prevalence of detectable heartworm infection within the specified population. However, given the known limitations of antigen testing in cats, particularly in cases of low parasite burden or immature infections, the absence of positive results should be interpreted with caution.

QUALITY CONTROL OF CANINE PLATELET CONCENTRATE PREPARED USING THE BUFFY COAT METHOD

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INTRODUCTION

The buffy coat (BC) method for canine platelet concentrate (PC) is widely used in human medicine but less reported in veterinary medicine. It involves initial high-speed centrifugation to isolate the buffy coat layer, followed by low-speed centrifugation to separate platelets from residual RBCs. Only limited canine studies exist, with no specific guidelines. Human AABB guidelines require $PLT > 5.5 \times 10^{10}/unit$.

This study describes the BC method for producing canine PC units and reports quality control results, comparing them with relevant current bibliography.

RESULTS

n=646	All units presented POSITIVE SWIRLING
Mean (SD) results	Volume 48 (±5) mL
	PLT $3.6 \times 10^{10}/unit$ ($\pm 1 \times 10^{10}/unit$)
	WBC $0.08 \times 10^3/\mu L$ ($\pm 0.16 \times 10^3/\mu L$)
	RBC $0.06 \times 10^6/\mu L$ ($\pm 0.11 \times 10^6/\mu L$)
Blood Culture	4 units with positive result (0.59%) Identified microorganisms: <i>Staphylococcus pseudintermedius</i> , <i>Streptococcus canis</i> , <i>Streptococcus dysgalactiae</i> , <i>Pasteurella canis</i> , <i>Streptococcus halichoeri</i> .

CONCLUSION

The BC method produces safe canine PC with low bacterial contamination and acceptable residual WBC and RBC levels, indicating improved leukoreduction and RBC removal compared to Hoareau *et al.* (2014).

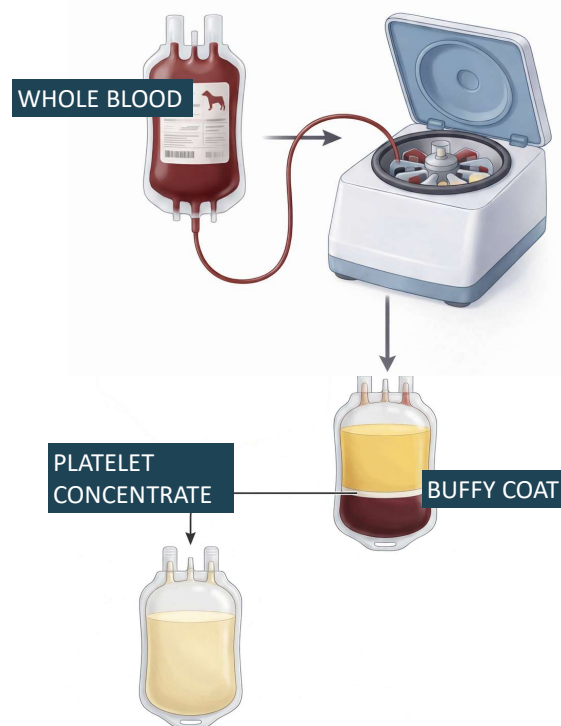
Platelet counts were higher than Hoareau *et al.* but below human standards (AABB), emphasizing the need for canine-specific guidelines that reflect the difficulty of achieving human standards.

METHODOLOGY

DATA 646 Platelet Units
Retrospective study
Feb 2025 - Jan 2026

DONORS Healthy
Mixed breed
1-8 yrs
>20 kg
Vaccinated/
dewormed
Never transfused
PCR/ELISA negative

BC METHOD 1st High-speed centrifugation (to isolate BF)
2485g, 17 min, acc 210s, dec 720s
2nd BC collection
3rd Low-speed centrifugation (to remove RBC)
100g, 10 min, acc 30s, dec 300s



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